

TEM • SAMPLE PREP • MICROPROBES • NMR • MASS SPEC

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## Solids for fun and games

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## Setup is Important!

- It is easier to get no signal than to get a signal in solids
  - You can guess at parameters for liquids NMR and get data. It may be poor but it will be data.
  - If you do this for solids you will get exactly what you deserve: NOISE!

## Things to know before you start:

- **Know your chemistry:**
  - **How was the sample prepared?**
    - Can result in inconsistent results
  - **What is the proposed structure?**
    - Can greatly affect NMR parameters
    - Molecular weight?
  - **What is the density?**
    - Can destroy the probe!
  - **What nuclei do you want to observe?**
    - Is the machine capable and will the probe tune?

## Example from customer:

- **Customer e-mailed and stated they:**
  - Ran Adamantane standard > great signals
  - Ran compound of interest > NO signal
  - Reran Adamantane standard > great signals
- **What is wrong?**
  - My first question: What is the unknown chemical?
  - Customer stated: Salicylic acid



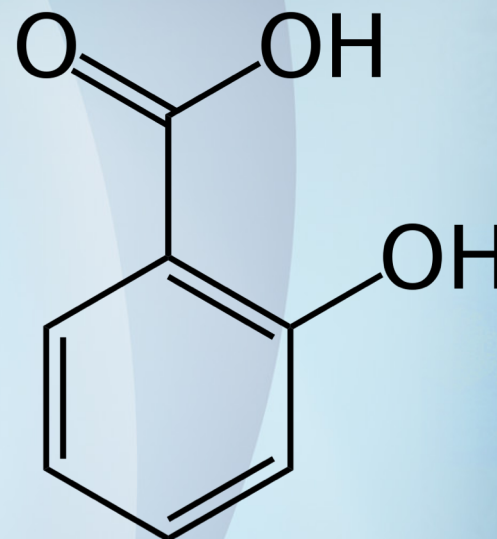
## Why No Signals?

- **What I did:**

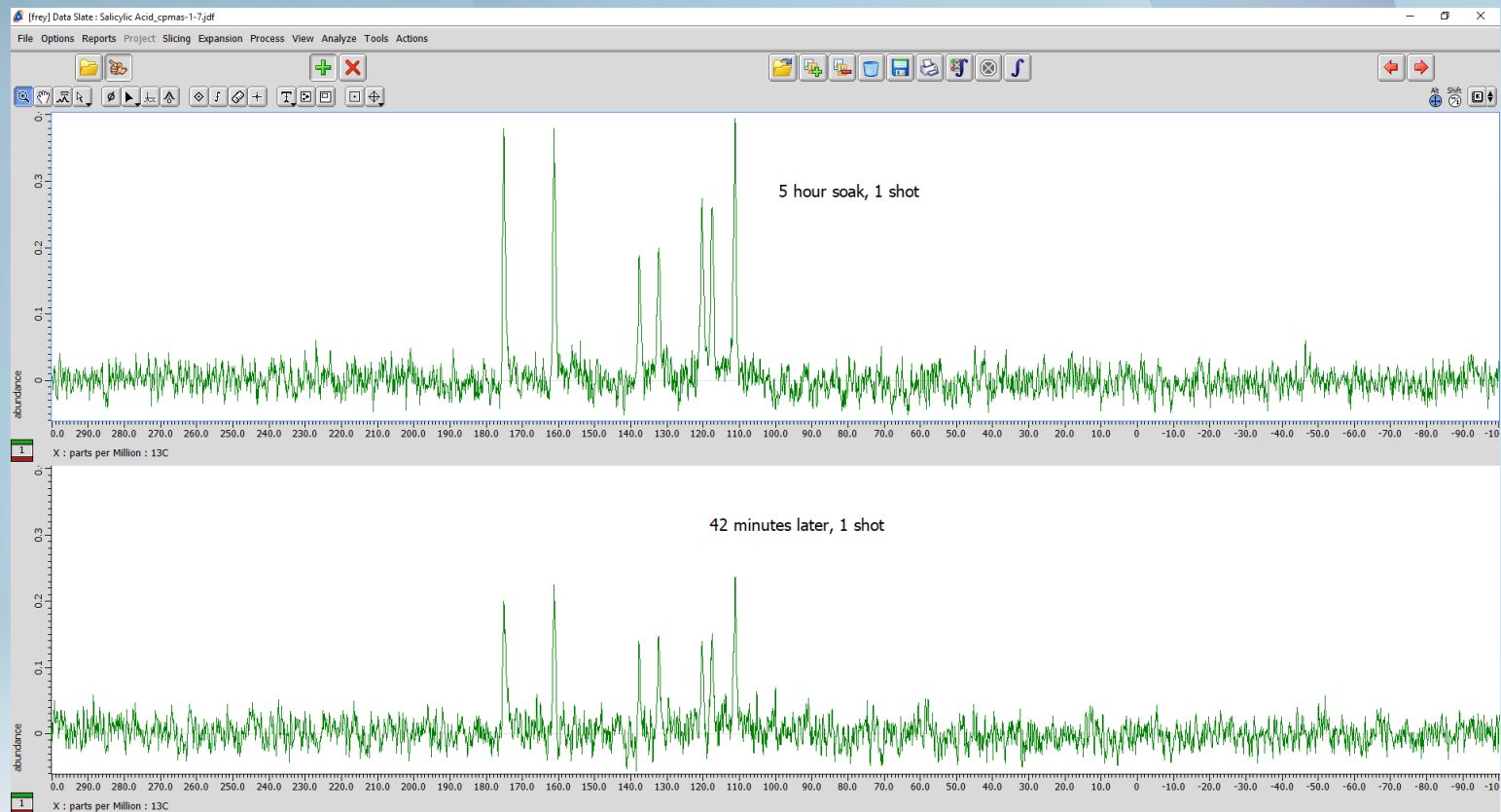
- I made up the sample and put it in the magnet
- Took 1 CP shot > No signal
- Let it soak in the magnet 5 hours
- Took 1 CP shot the next morning > Great Signal!!
- Waited 1 hour
- Took 1 CP shot > poor signal

- **Result**

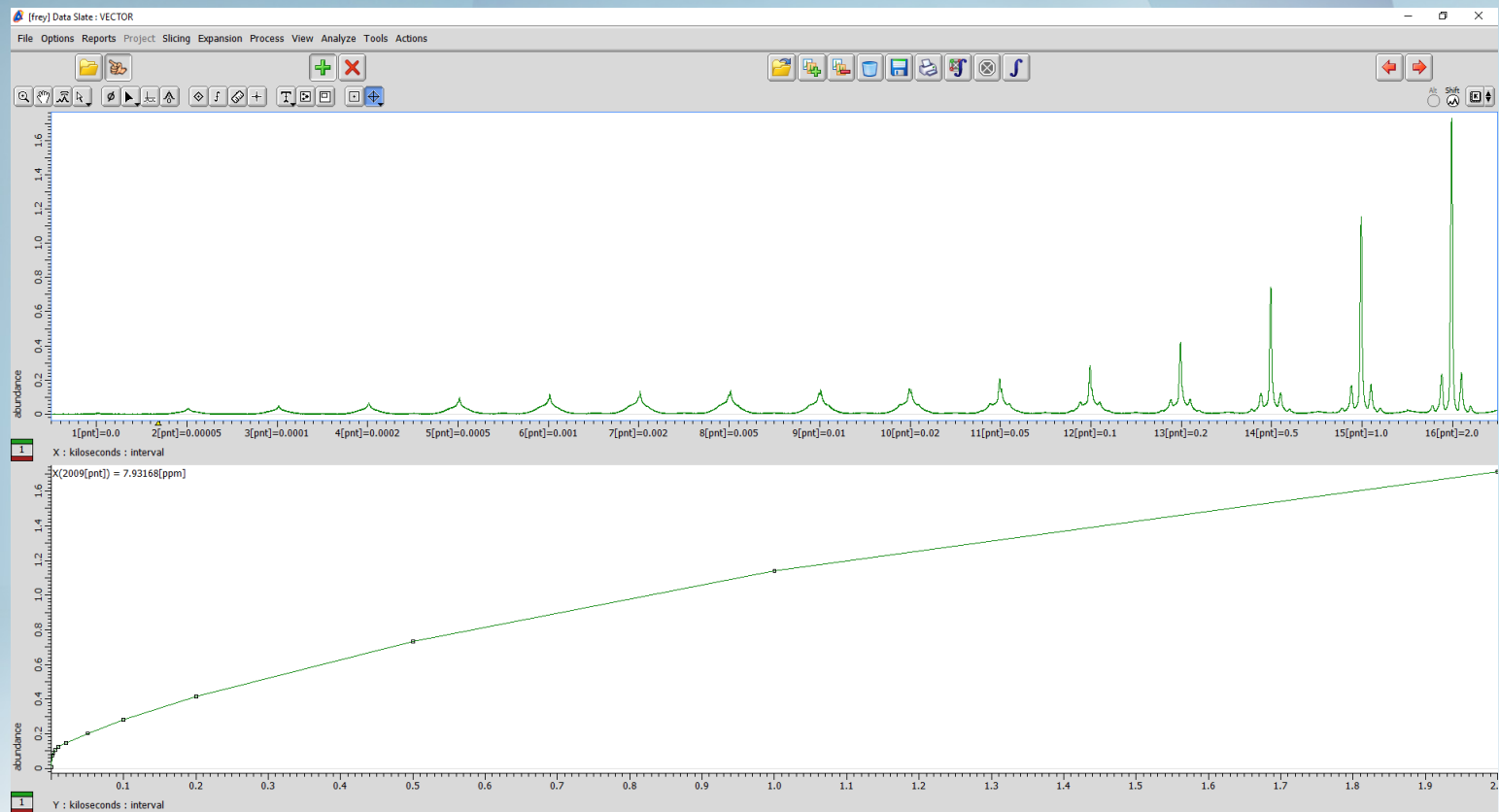
- The  $^1\text{H}$  T1 is very very long (hours!).
- You need internal motion to bring down Proton T1
  - Includes Methyl Groups, Phenyl Rings, soft materials



# Know your chemistry



# Proton Saturation Revocery



## Packing and Spinning Samples

- **Pack loosely**
  - Compressing the sample is usually not necessary
- **Put sample in vial and label the vial!!**
  - You cannot write the sample name on the rotor
- **Test in Bench Spinner**
  - It is less expensive to repair the bench spinner than the MAS NMR probe



## Experiments and sample parameters for setup

- KBr – Magic Angle
- $^1\text{H}$  90 Array – Proton Power
- CP Array – Match condition
- Contact Array – Maximum Signal
- Ramp Array – Equalize CP to all sites
  
- Examples are for 3.2mm HXMAS probes
  
- Typical spinning speeds for  $^{13}\text{C}$  = ~100 to 125 ppm
  - $^{13}\text{C}$  1<sup>st</sup> order spinning sidebands are minimal
  - 400 = 10 to 12.5 kHz
  - 500 = 12.5 to 15 kHz
  - 600 = 15 to 20 kHz

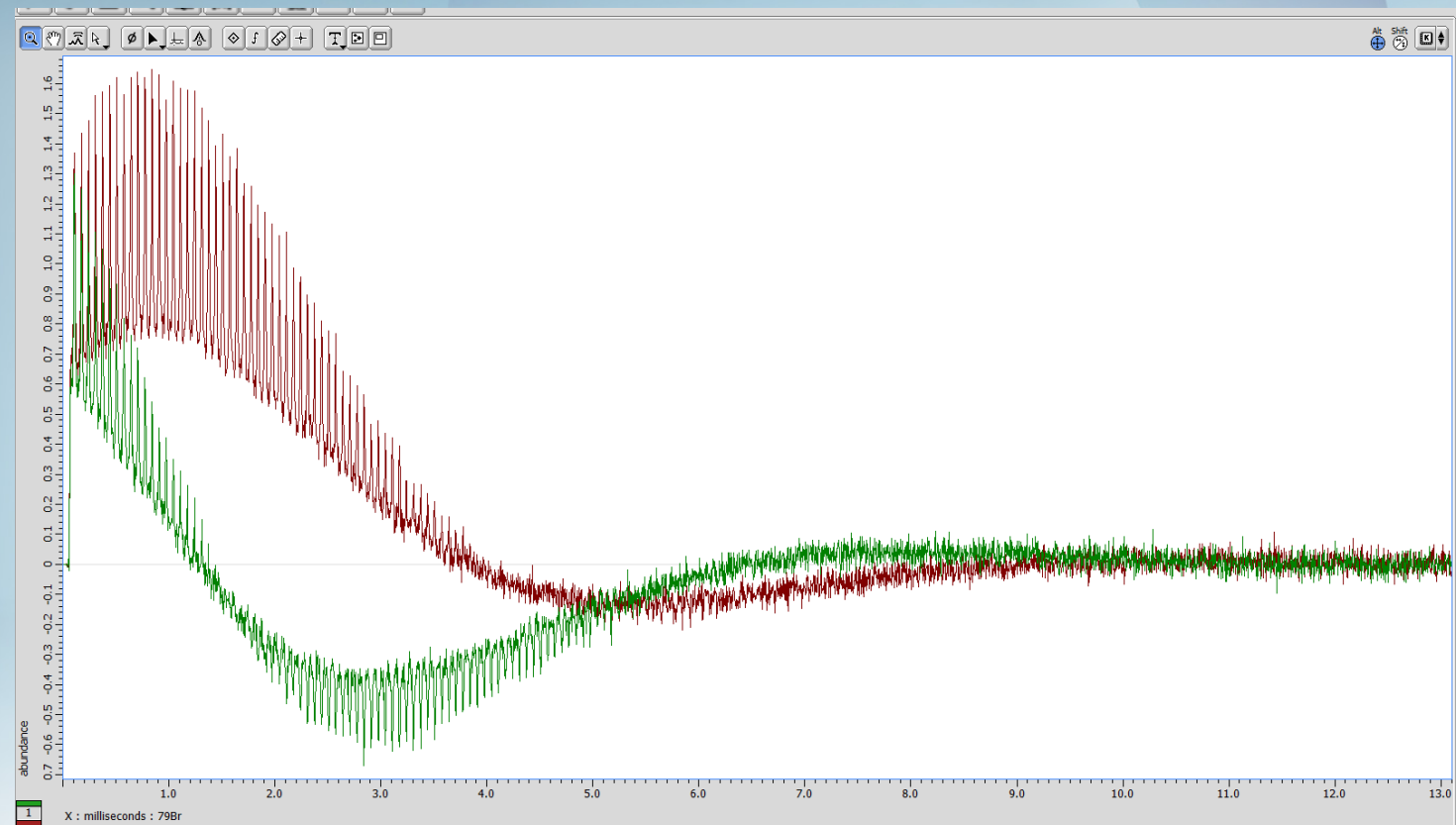
## Standard $^{13}\text{C}$ Setup Sample - AGK

- **Mixture of:**
  - Adamantane: 227mg
  - Glycine: 500mg
  - KBr: 291mg
- **Equal molar in  $^{13}\text{C}$  carbon sites**
- **Saves expense and repacking rotors for multiple standards**
- **Works for:**
  - Setting Magic Angle
  - Setting CP conditions
  - Checking Proton Decoupling
  - Chemical Shift Reference for  $^{13}\text{C}$ , Adamantane peak at 37.77 ppm.
- **Pure Adamantane will have sharper lines**

## KBr Magic Angle Adjustment

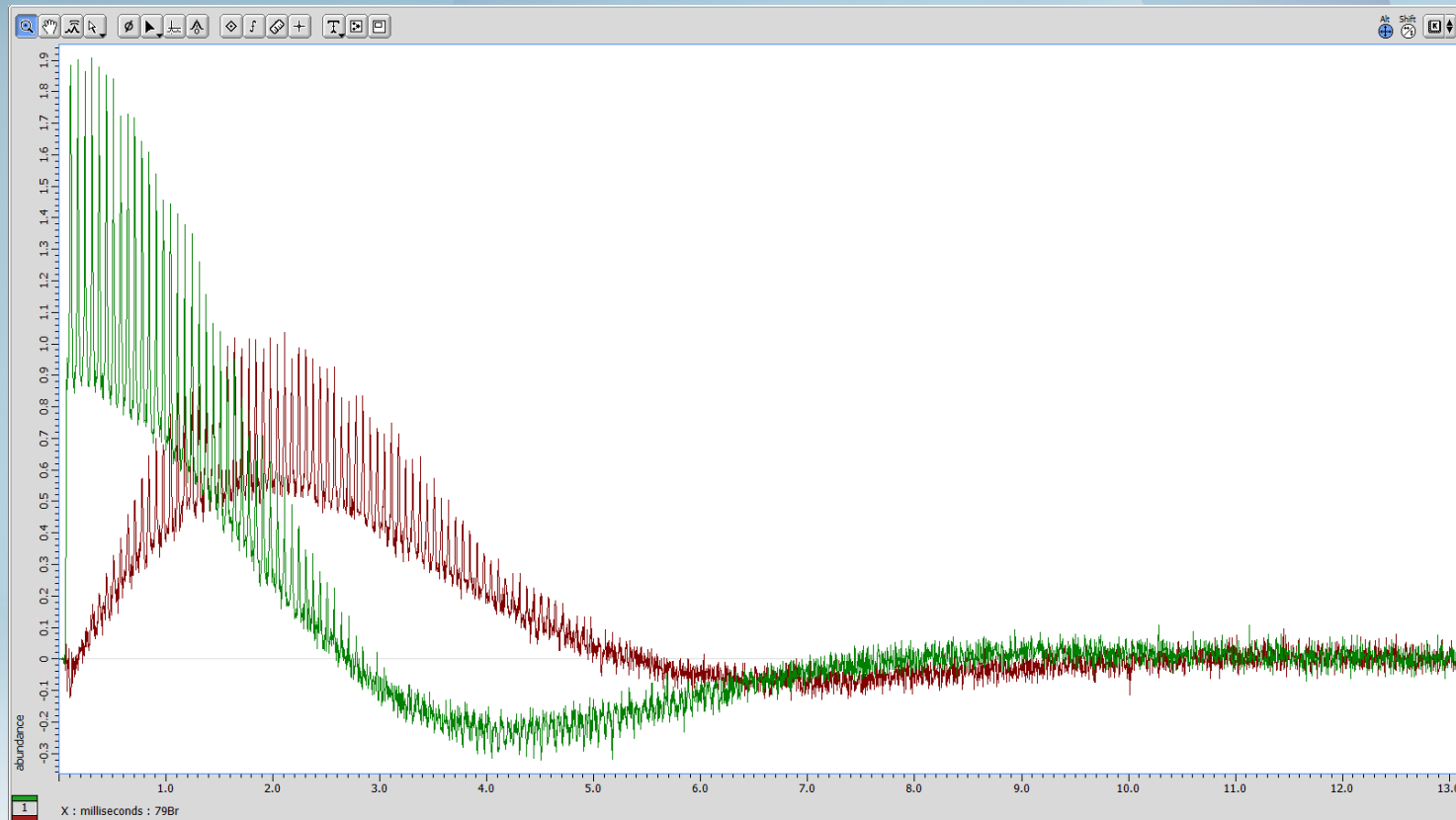
- Use mas\_adjust\_kbr.jxp experiment
- Load and start the experiment
  - If there are no parameters for  $^{79}\text{Br}$  use  $^{13}\text{C}$  values
  - Tune probe
- Set up monitor:
  - Process DC balance
  - Press Shift-2 to display imaginary data (red)
  - Set phase point ( $\Phi$  p) to 0%
  - Adjust PO to get Imaginary FID (red) to start at 0
  - Adjust P1 to remove oscillations from FID

# KBr Magic Angle Adjustment 1 – No Adjustment

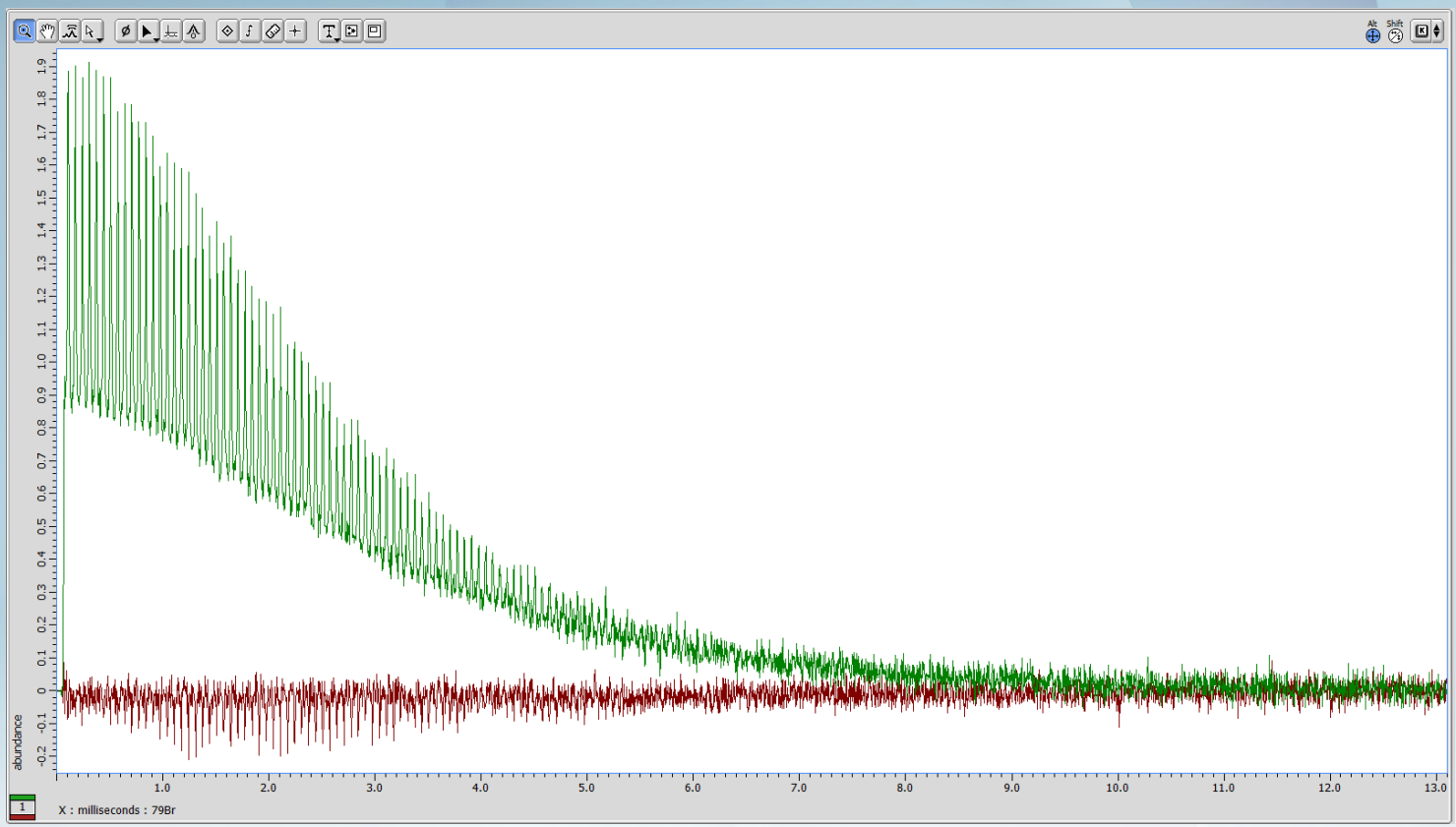




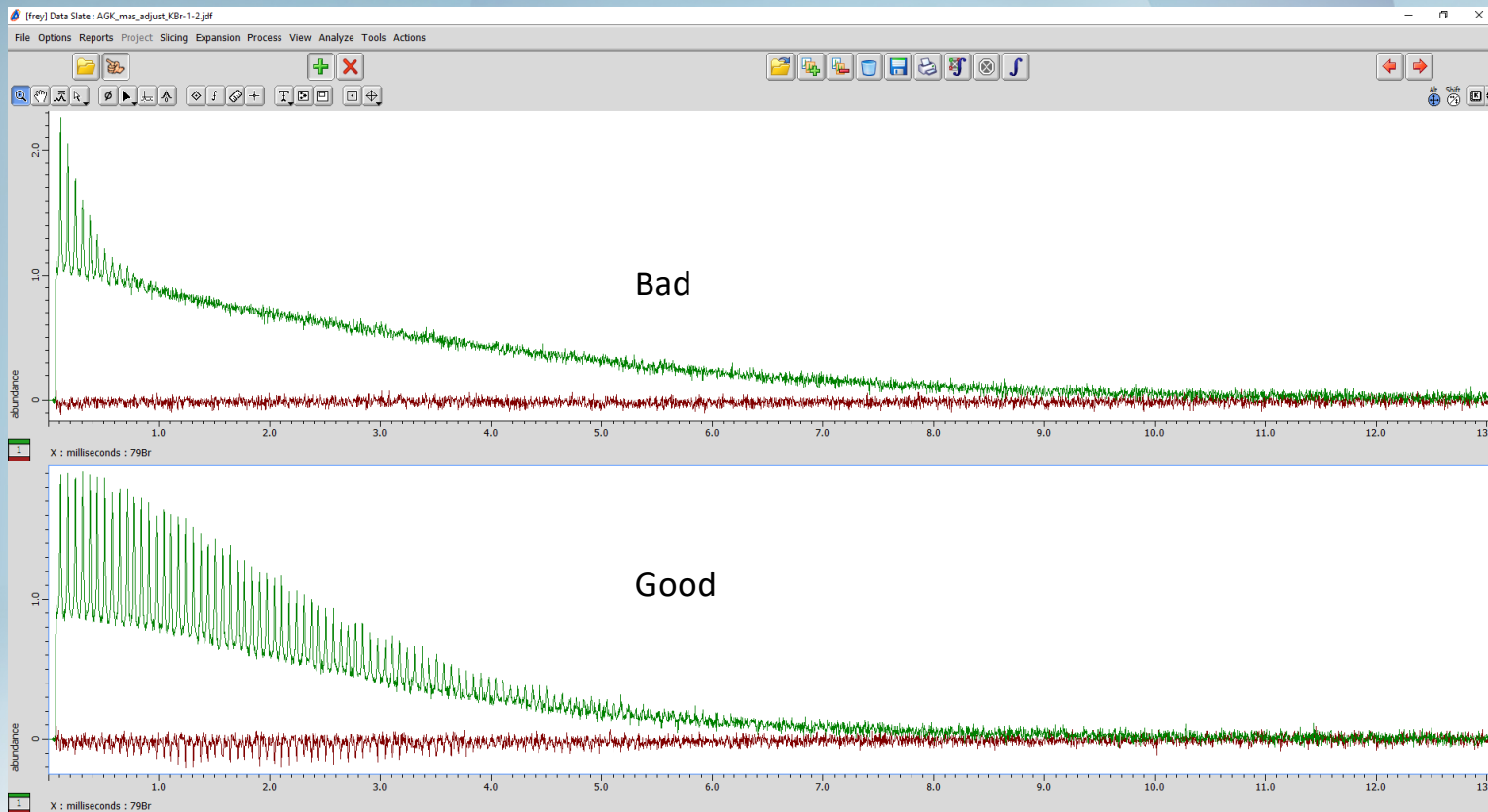
# KBr Magic Angle Adjustment 2 – After P0 Adjustment



# KBr Magic Angle Adjustment 3 – After P1 Adjustment



# KBr Magic Angle Adjustment 4 – Angle Adjustment

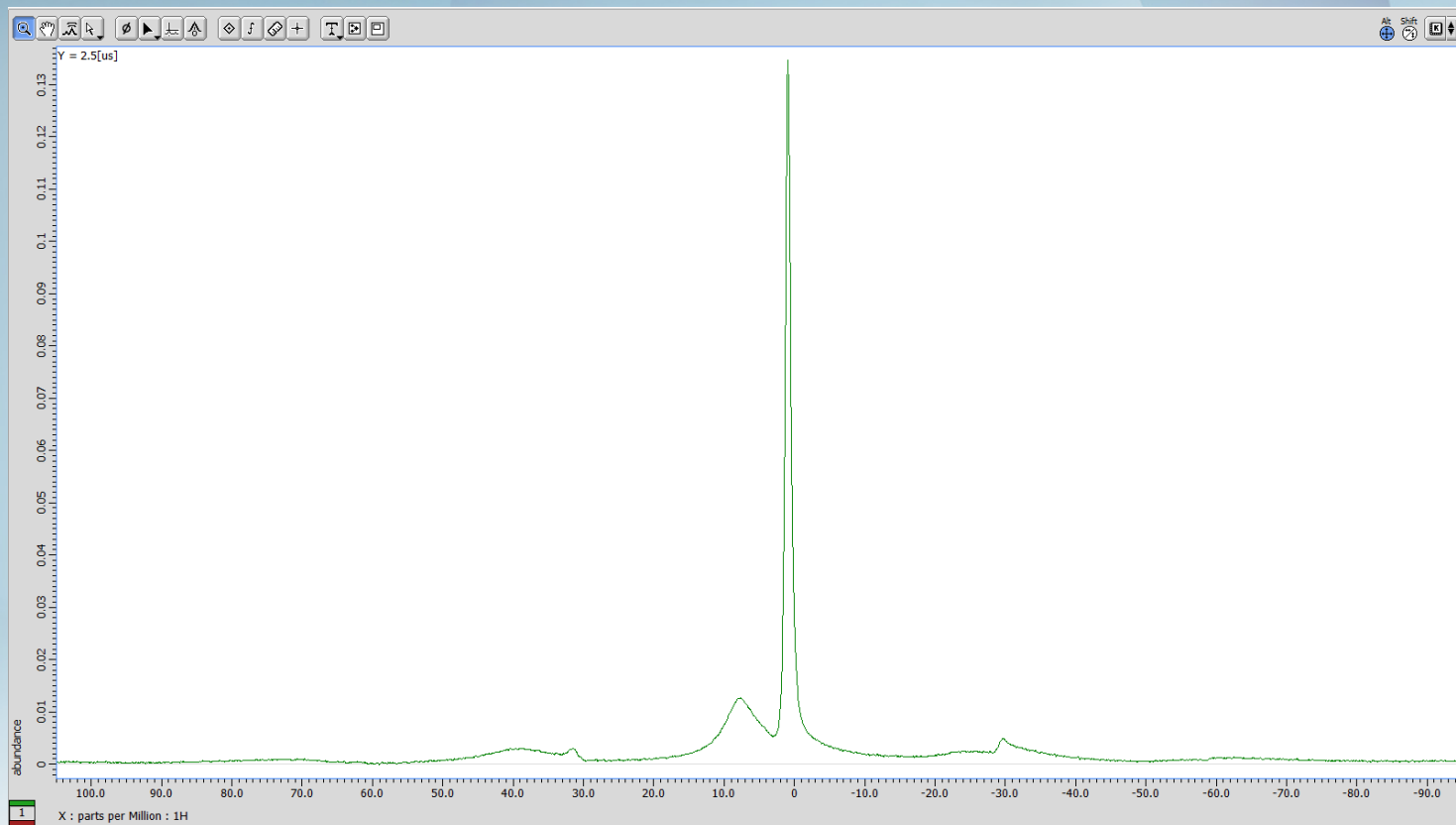


## **$^1\text{H}$ 90 Check**

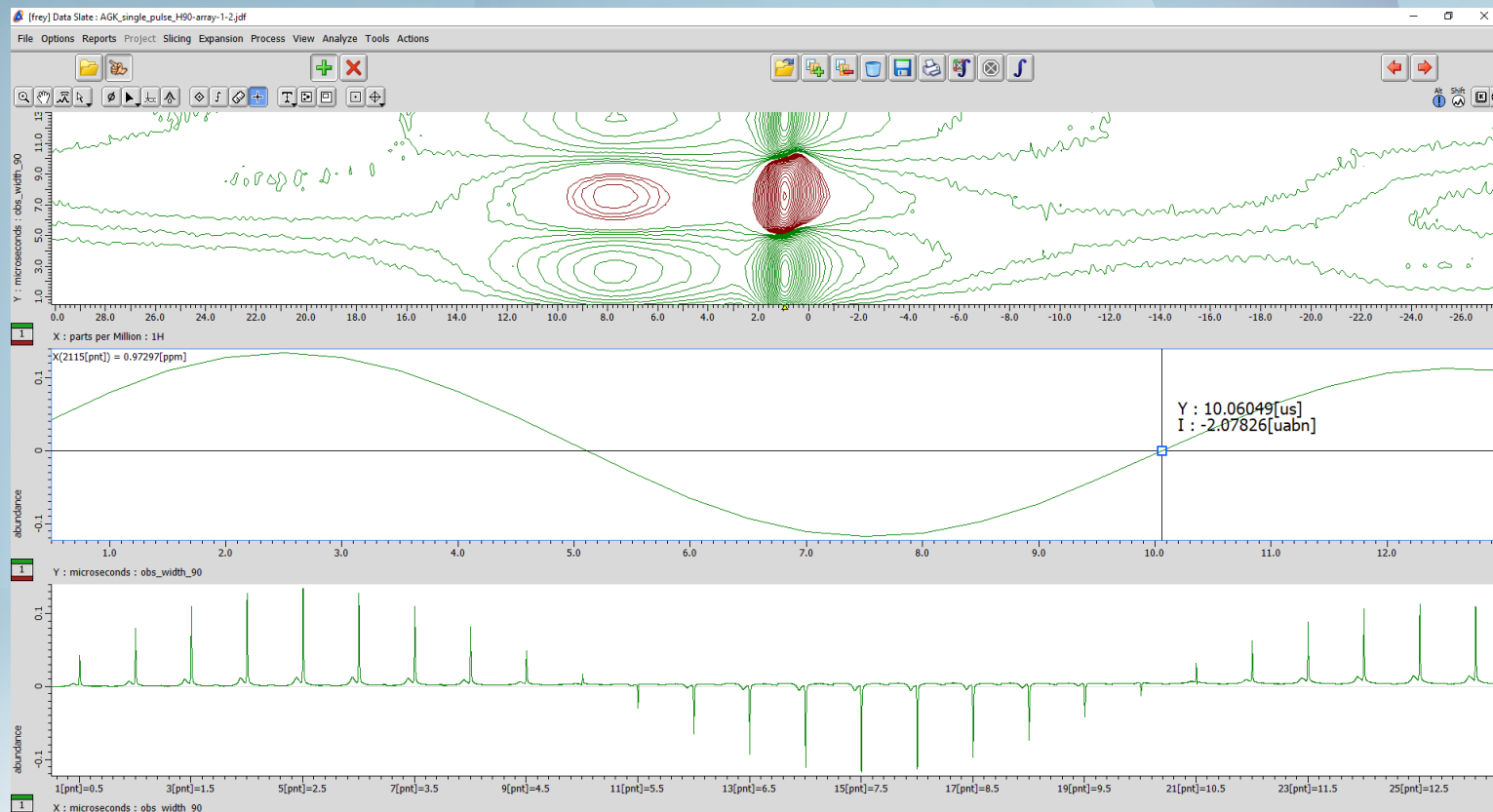
- Use single\_pulse\_solid.jxp
- Run just like  $^1\text{H}$  90 check in liquids
- To make sure  $^1\text{H}$  decoupling is working properly
  - Power levels are critical in solids



# $^1\text{H}$ spectrum of AGK



# $^1\text{H}$ 90 Array results – H90 = 2.5 us



## Setup CPMAS

- CP array to determine CP power
- Contact Array to determine best contact time
- Ramp Array to maximize all signals
- Use CPMAS cpmas.jxp

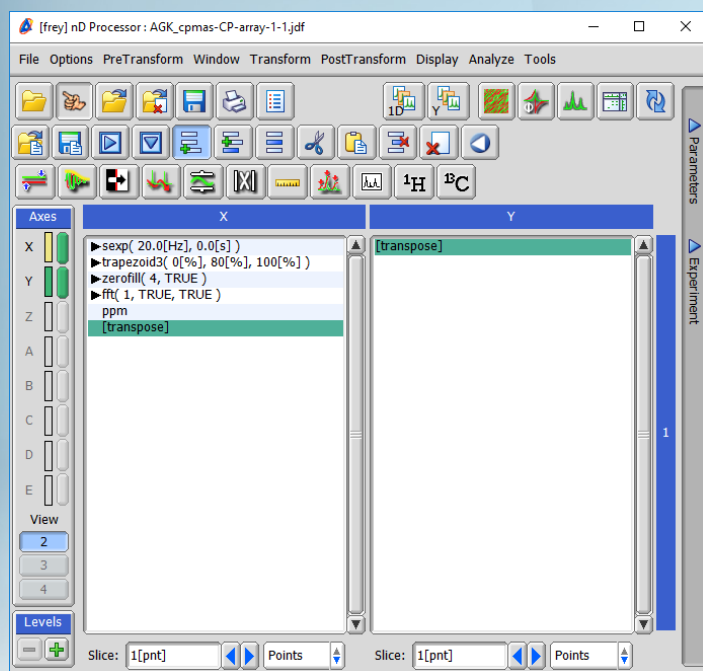
## Setup CP array

- Scans = 1
- obs\_shape\_cp = constant\_cp
- contact\_time = 5 ms
- irr\_amp\_cp = 50%
- obs\_amp\_cp = y\_acq 10[%]..80[%] : 1[%]

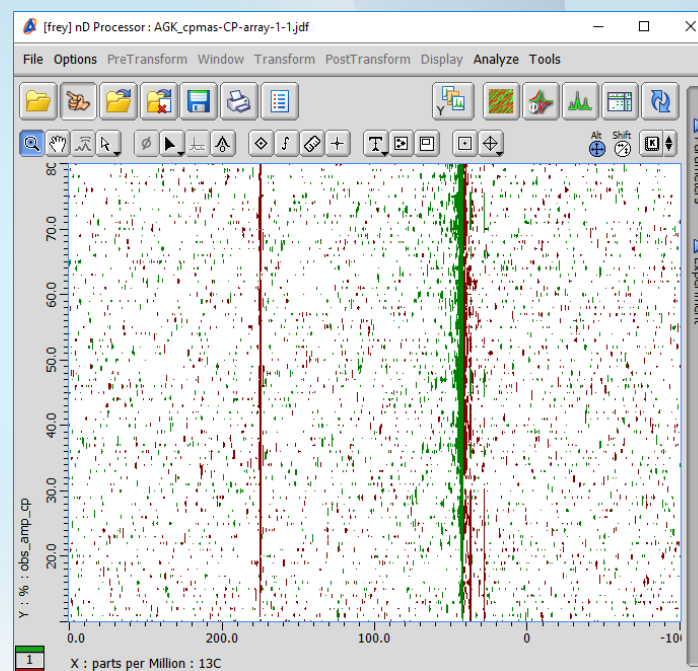


# CP -Array

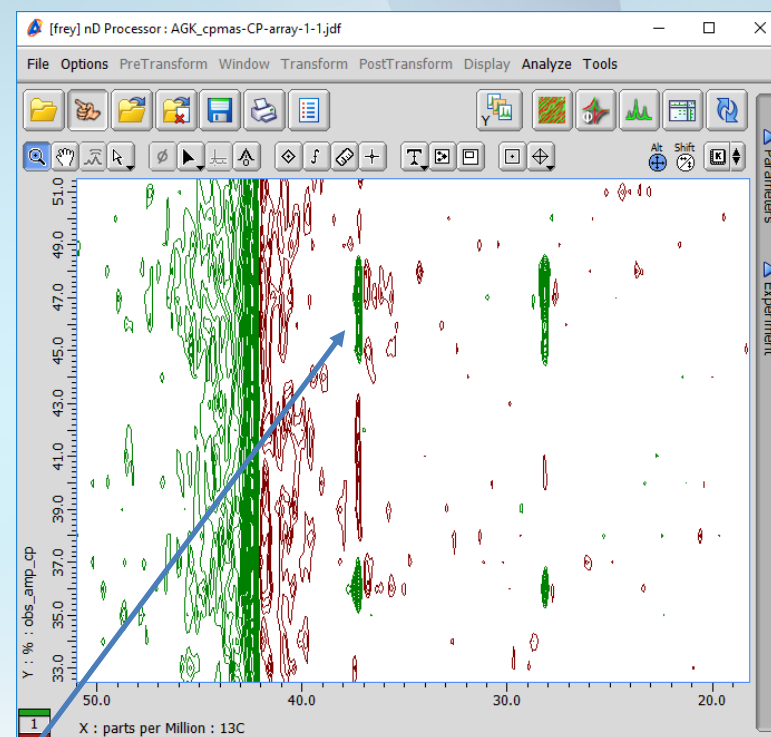
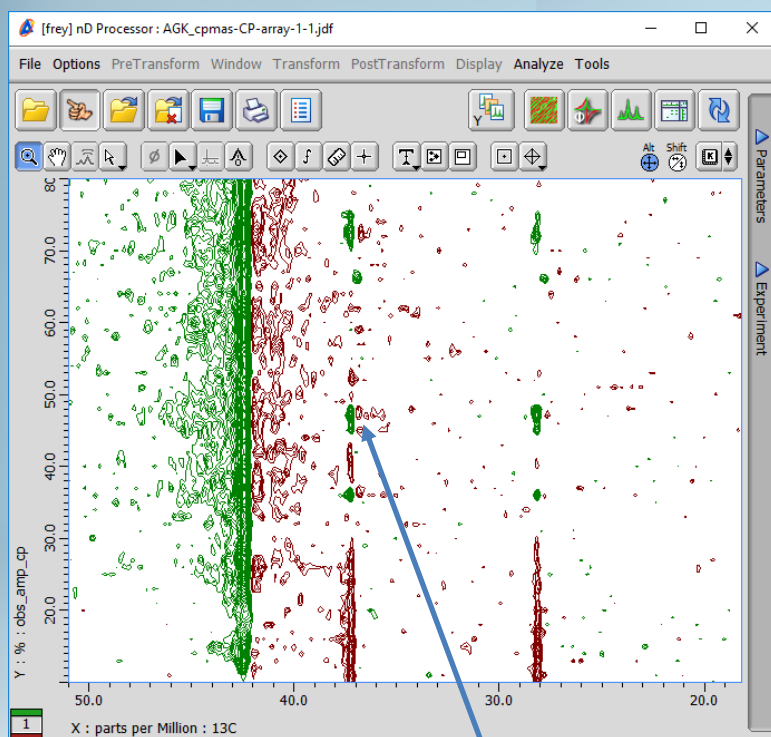
## Processing List



## Un-Phased Result

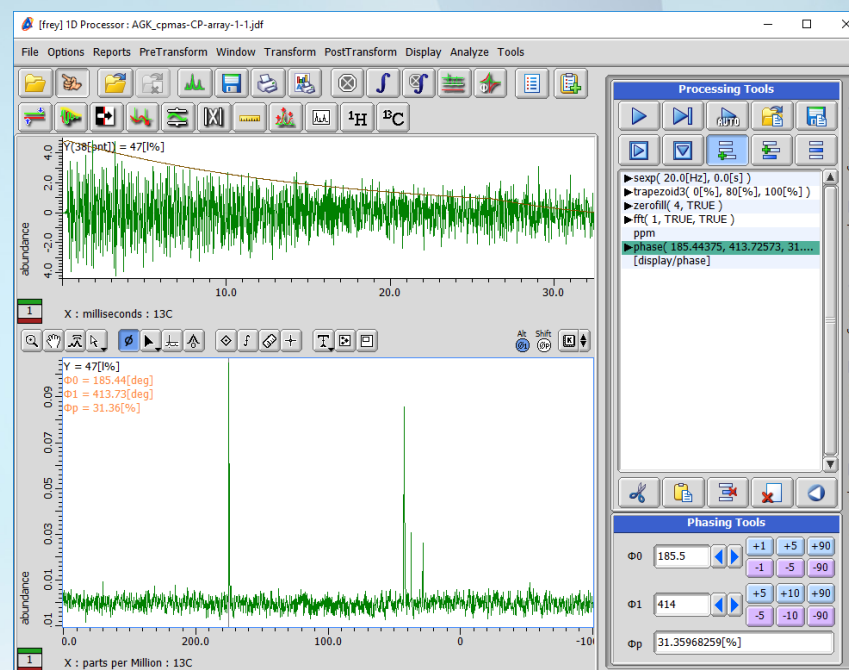
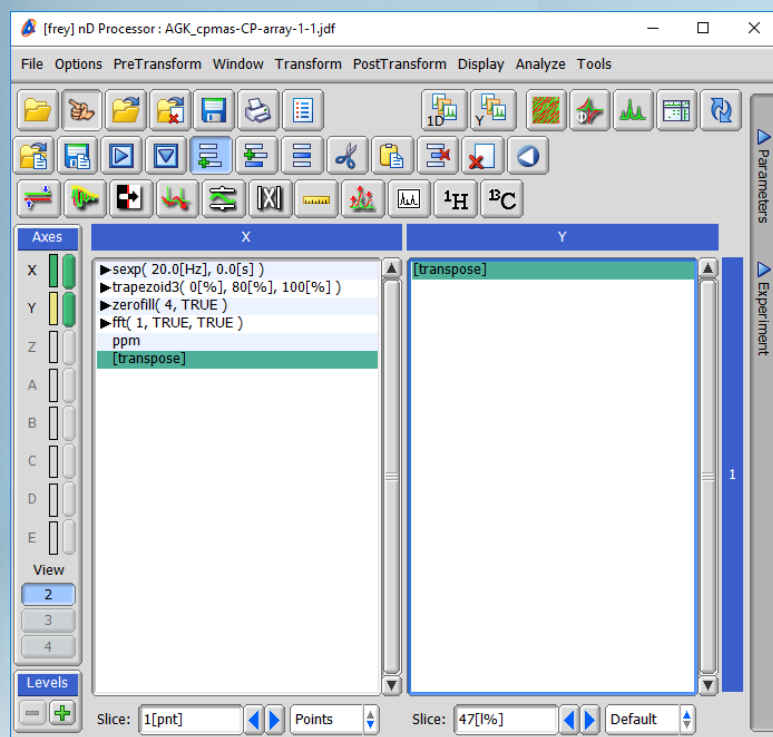


# Zoom on Aliphatic region to find Adamantane peaks

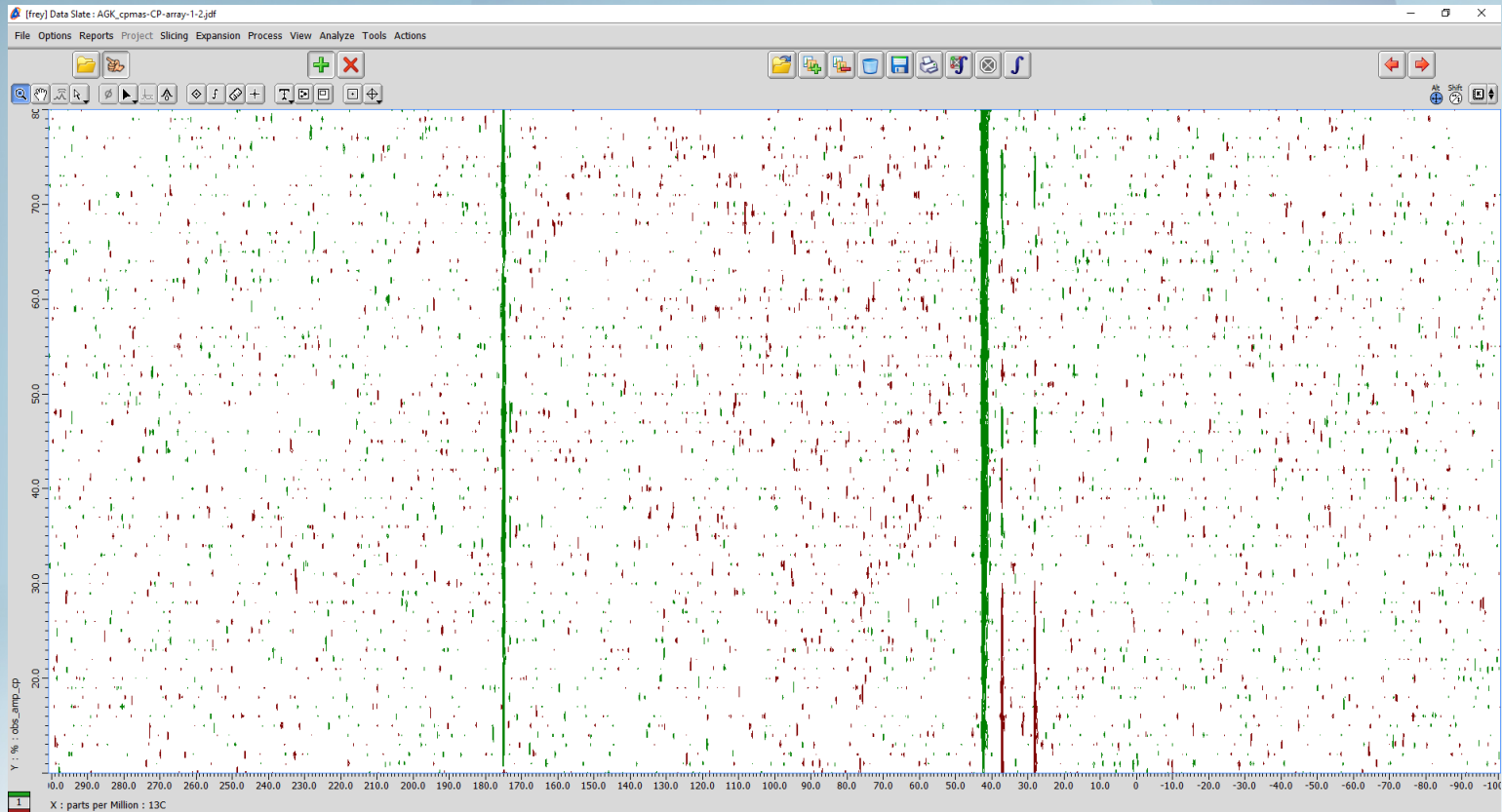


Adamantane peak

# Pick slice at 47% to phase



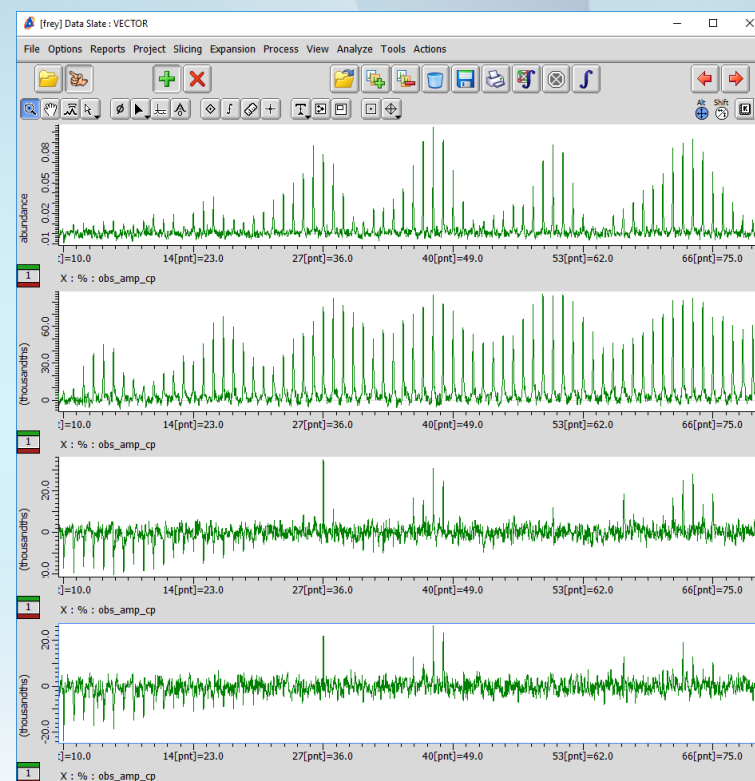
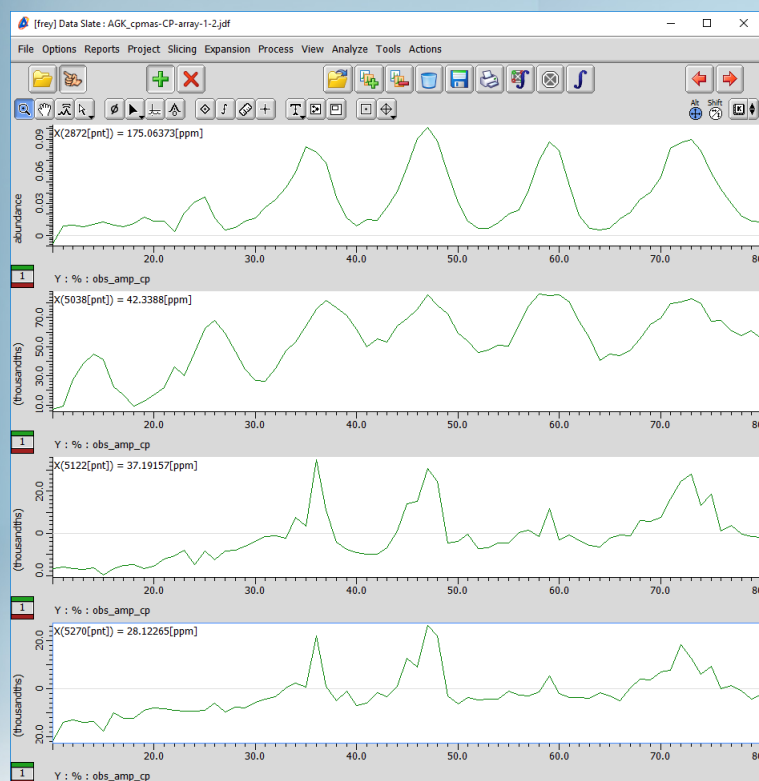
# Process the data to Data Slate





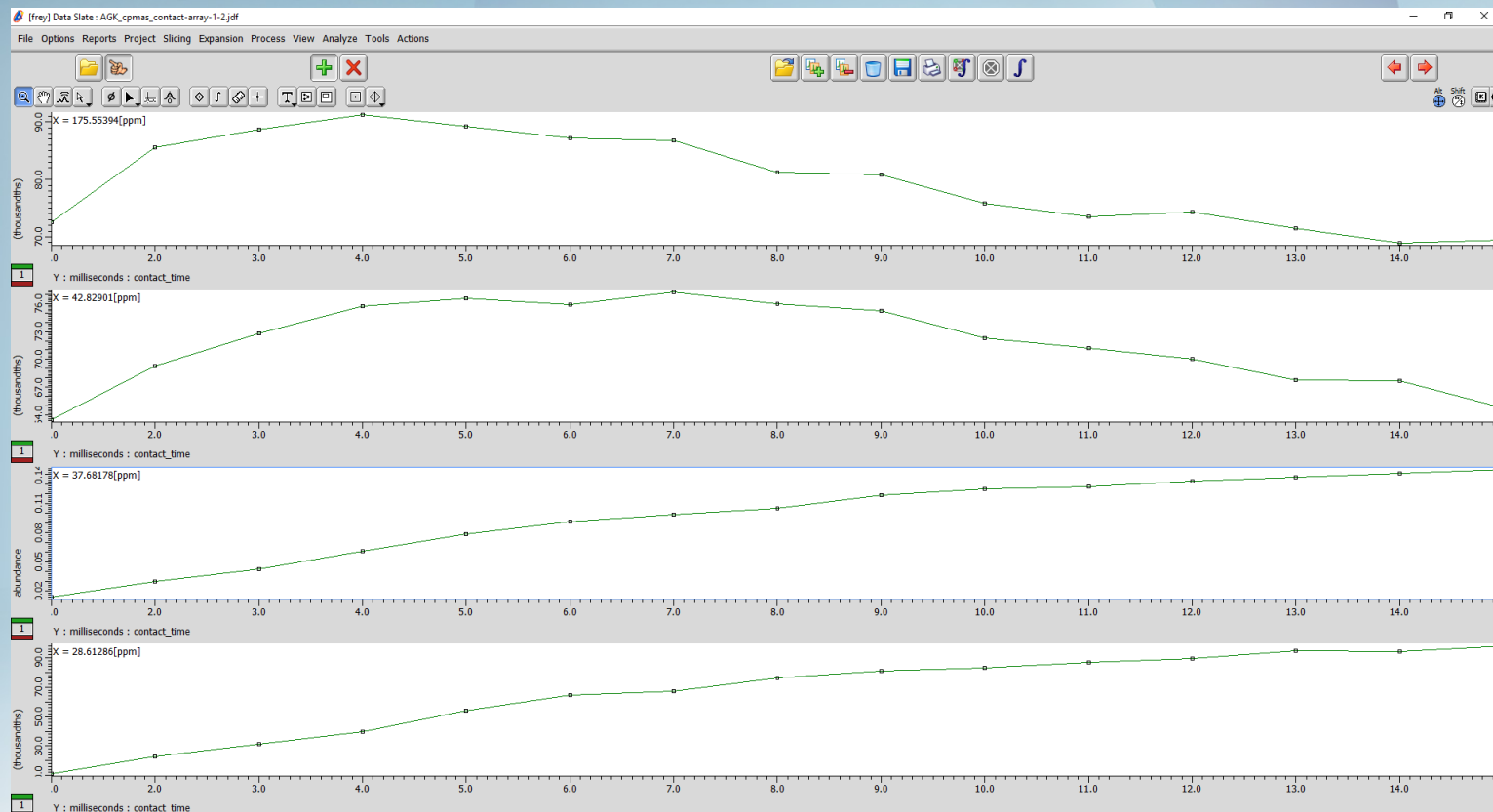
# Pick slices or linearize at peak positions

Update obs\_amp\_cp to use maximum @ 47%

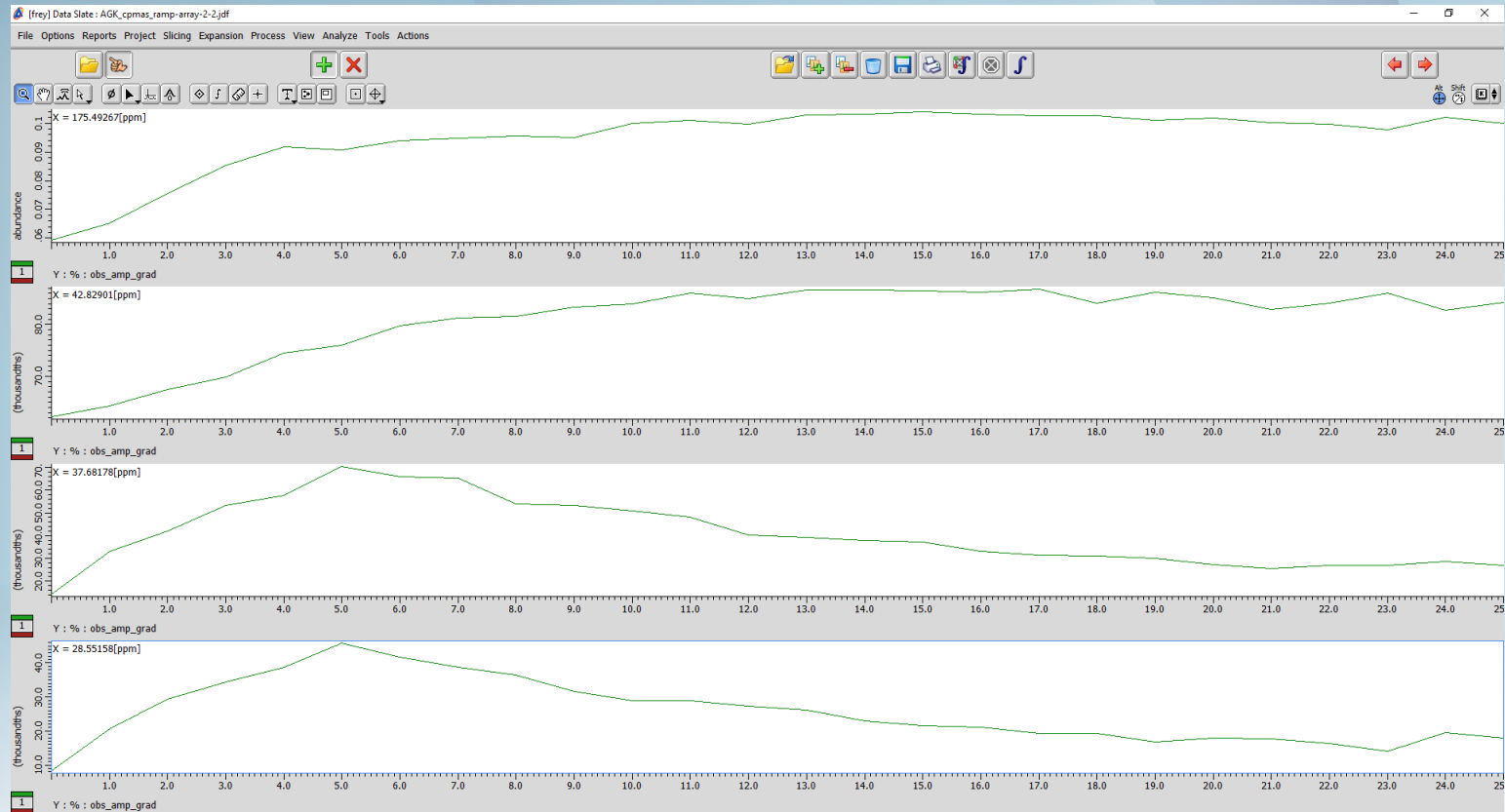




# Setup contact array = y\_acq {1[ms], 2[ms], 5[ms], 7[ms], 10[ms]}



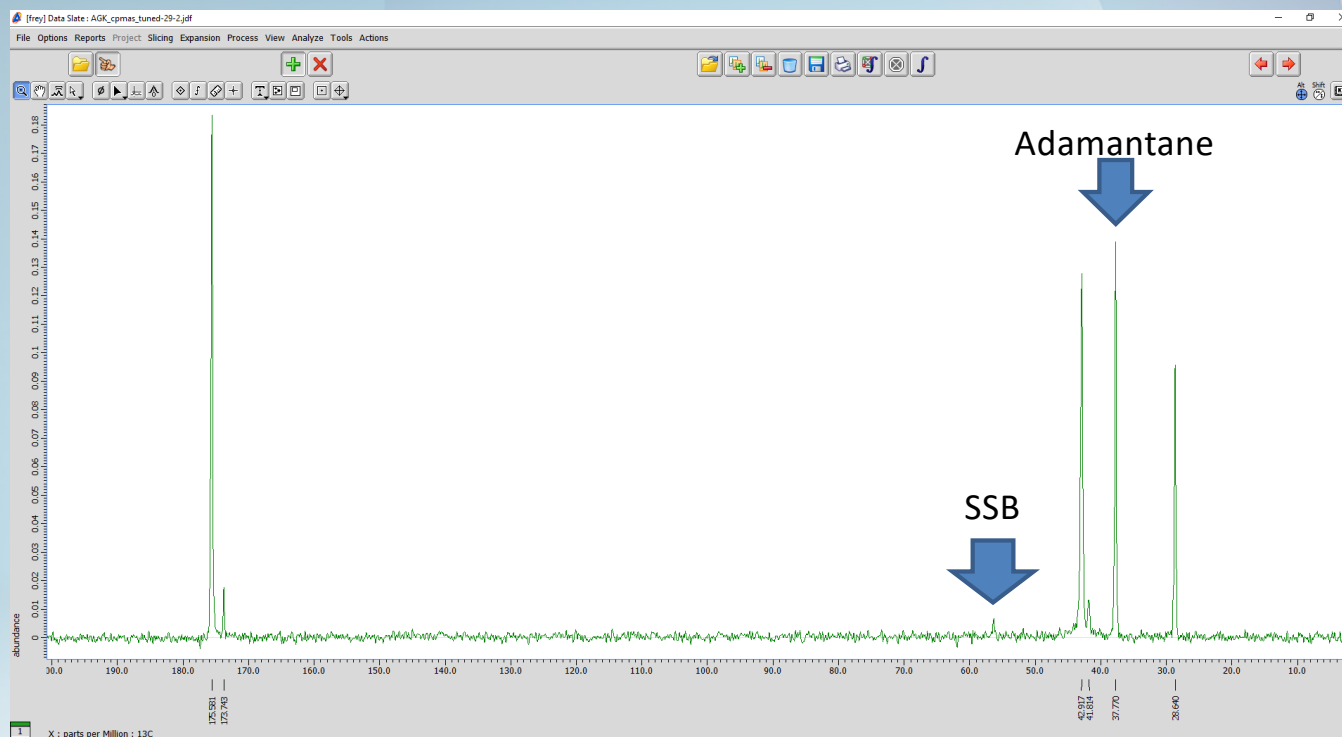
# Setup Ramp array = y\_acq 0[%]..20[%] : 1[%]



## CPMAS Tuned experiment

- Scans = 16
- obs\_shape\_cp = RAMP\_cp
- contact\_time = 4 ms
- irr\_amp\_cp = 50%
- obs\_amp\_cp = 47%
- obs\_amp\_grad = 5%

# AGK Tuned CPMAS spectrum



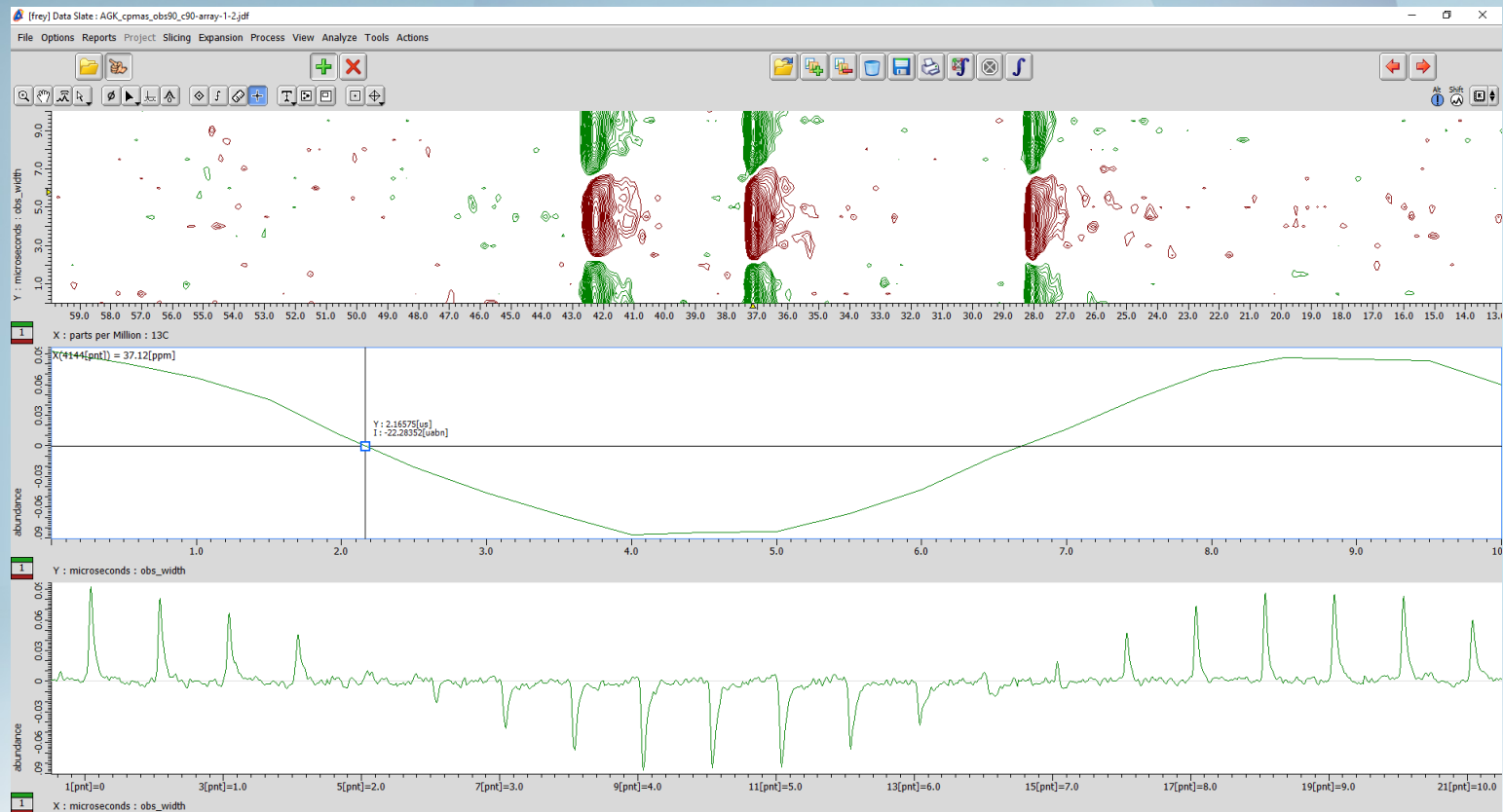
Adamantane peak is also chemical shift reference at 37.77 ppm  
Spinning speed 15 kHz

## **$^{13}\text{C}$ 90 via CPMAS**

- Use cpmas\_obs90.jxp experiment
- Set up standard CP conditions
- Array obs\_width = y\_acq 0[us]..10[us] : 0.5[us]



# $^{13}\text{C}$ 90 via CPMAS



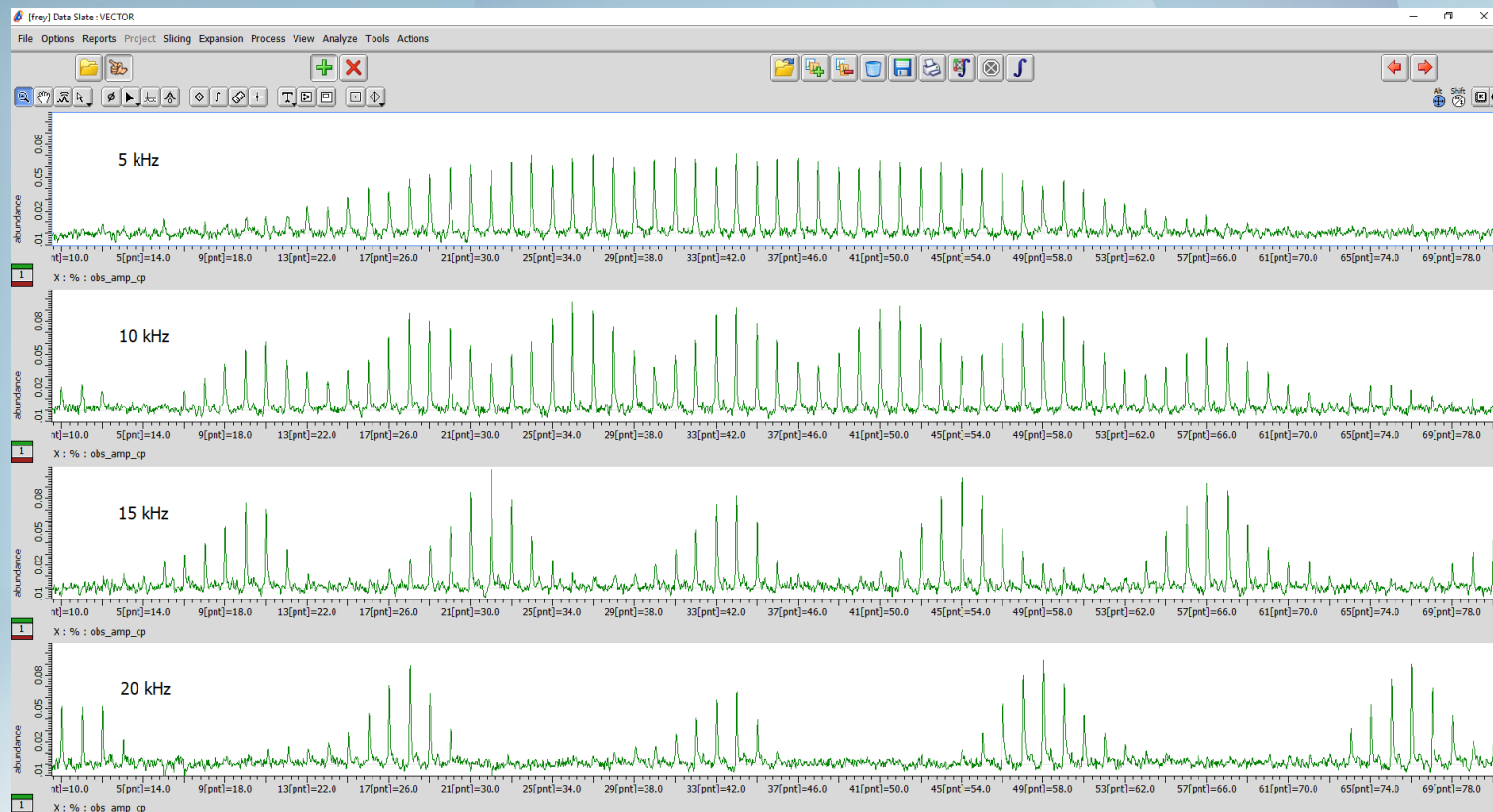
## Running Samples Standard Procedure

- Run tuned CPMAS of AGK before every sample
- Run sample
- Run tuned CPMAS of AGK after every sample
- This does 3 things:
  - Checks that the machine is operating correctly and the problem is the sample not the spectrometer
  - Provides chemical reference for all spectra.
  - Check if magic angle is drifting
- This only adds a few minutes, but ensures that the instrument is working properly before and after every sample

## Things to check

- If sensitivity on real sample seems low:
  - Run quick contact time array on real sample
  - Run a quick  $^1\text{H}$  T1
- Rerun the CP obs amp array if spinning speed changes

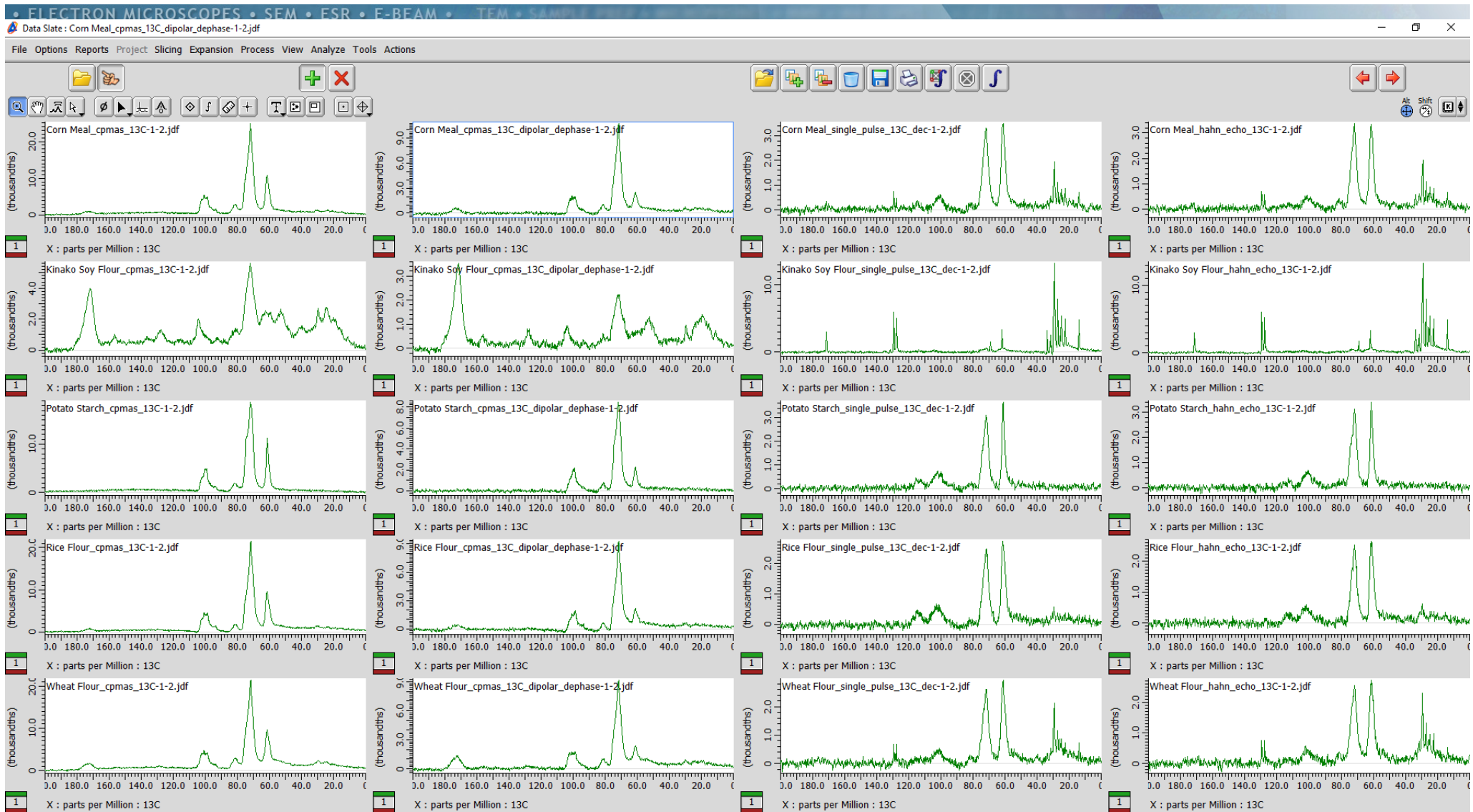
# CP Array vs Spinning Speed

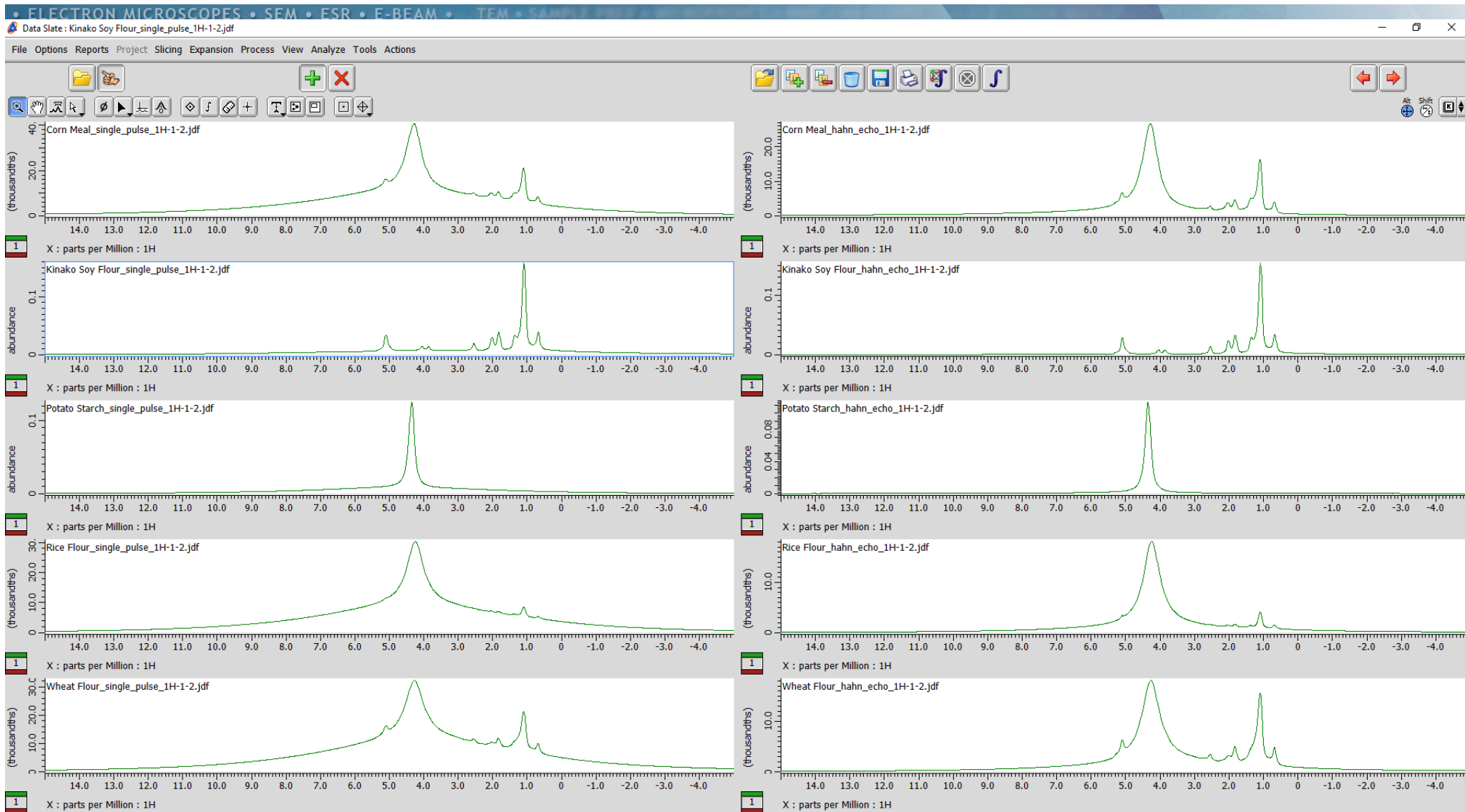


## Other Experiments to remember:

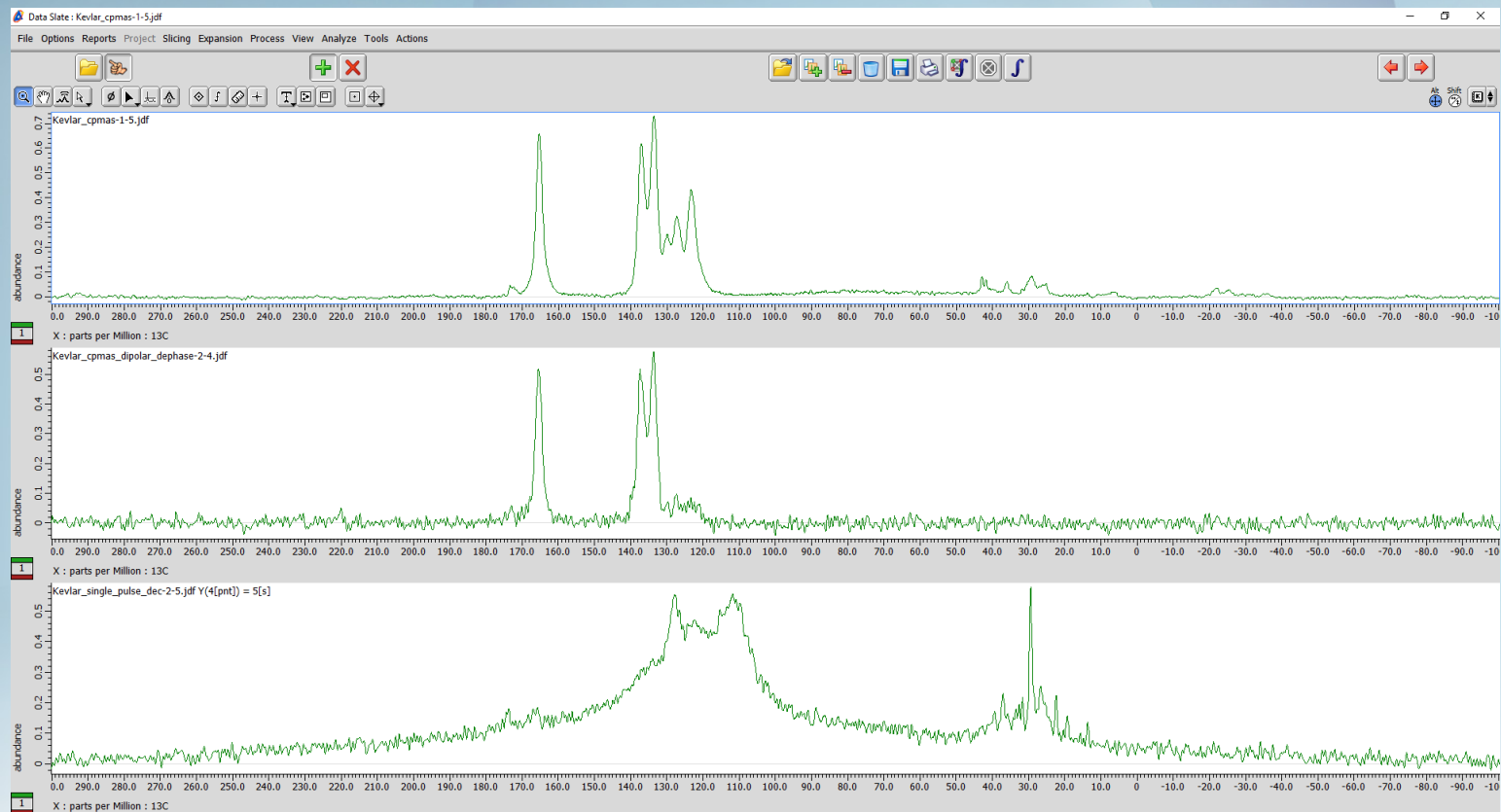
- $^1\text{H}$  Single Pulse with DEPTH
- $^1\text{H}$  Echo
- $^{13}\text{C}$  Single Pulse with DEPTH
- $^{13}\text{C}$  Echo
- $^{13}\text{C}$  CPMAS Dipolar Dephasing



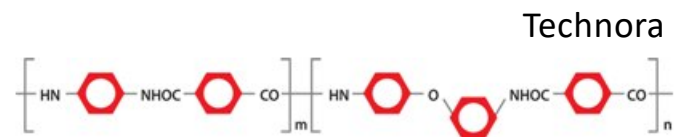
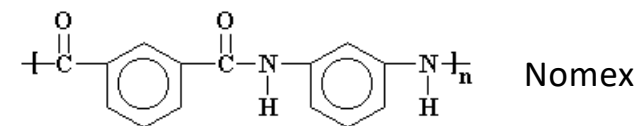
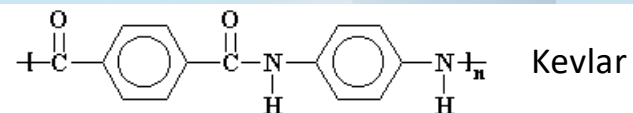
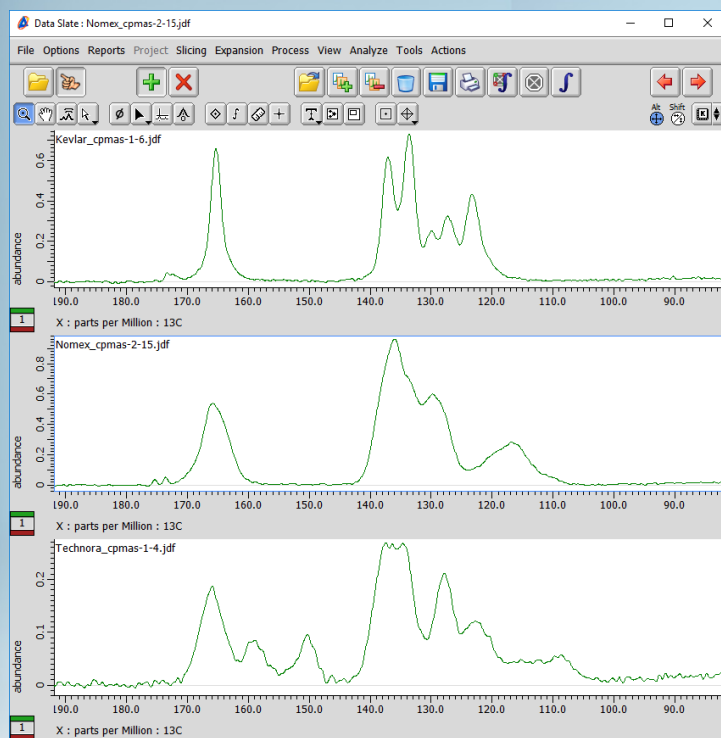




# Polymer example - Kevlar



# Polymer example – Aramid Fibers

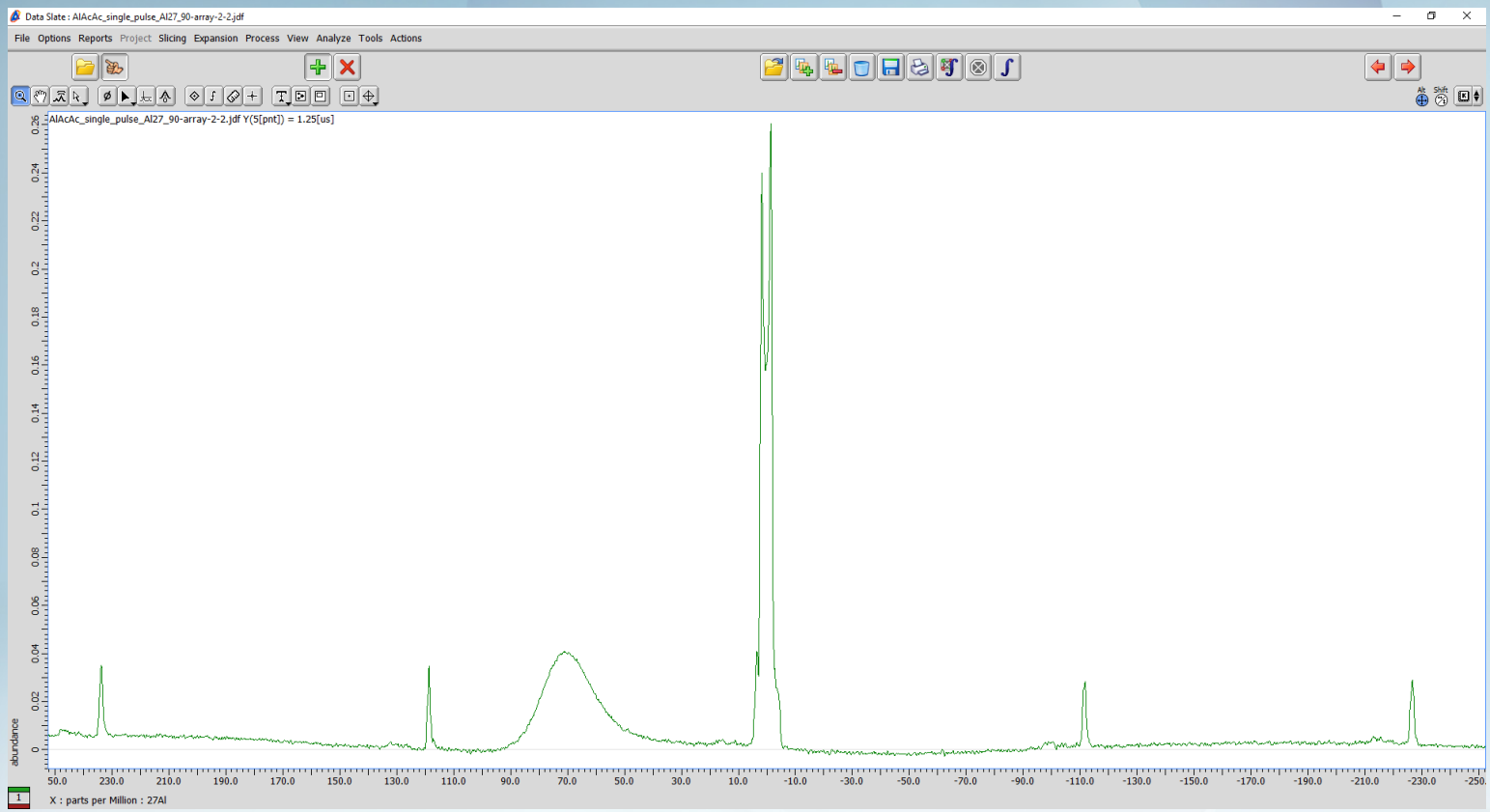


## 3Q MQMAS Split $t_1$ Echo with Fast Amplitude Modulation Pulse

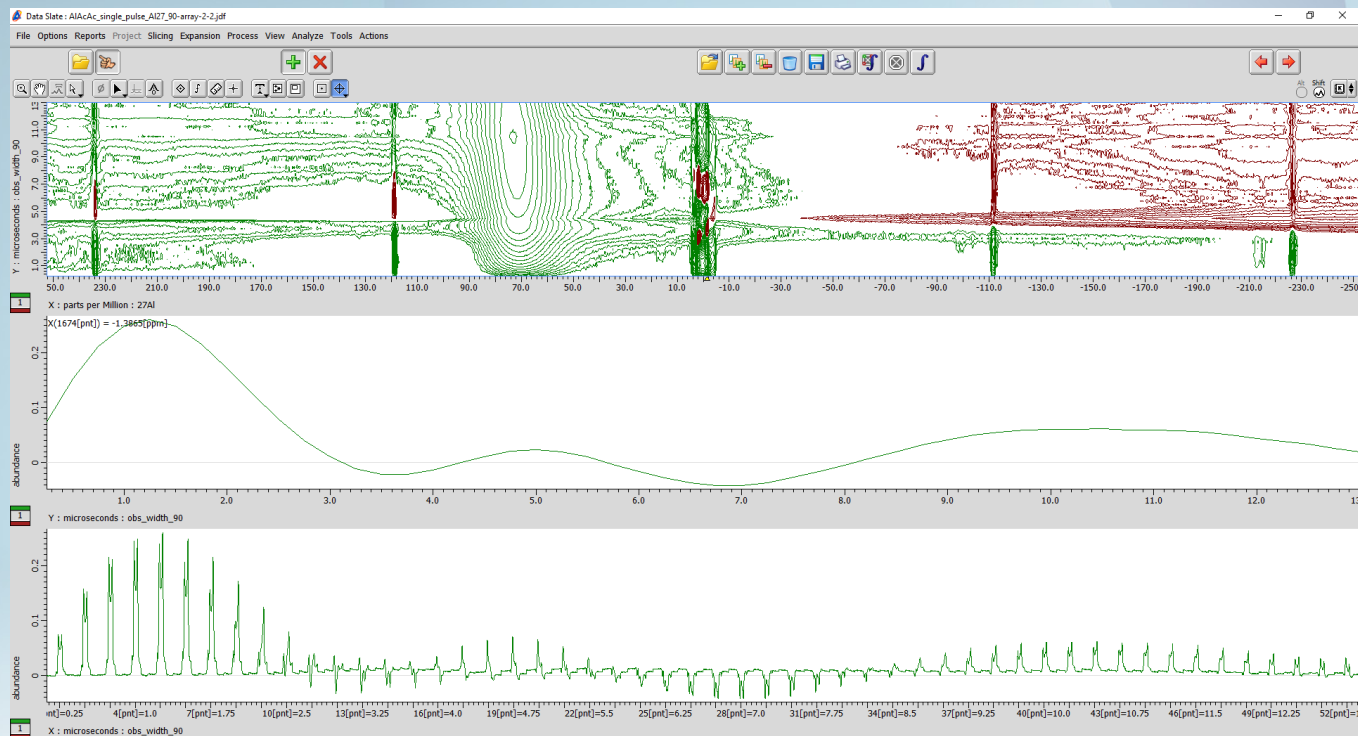
- 3 Quantum MQMAS Experiments:
  - mqmas-s3-3q\_fam\_split-t1\_echo.jxp for Spin 3/2
  - mqmas-s5-3q\_fam2\_split-t1\_echo.jxp for Spin 5/2
  - mqmas-s7-3q\_fam2\_split-t1\_echo.jxp for Spin 7/2
  - mqmas-s9-3q\_fam2\_split-t1\_echo.jxp for Spin 9/2
  - And setup versions
- Processing lists:
  - mqmas\_split-t1\_echo.list
  - mqmas\_split-t1\_echo\_check.list



# Easy test sample AlAcAc for $^{27}\text{Al}$ MQMAS

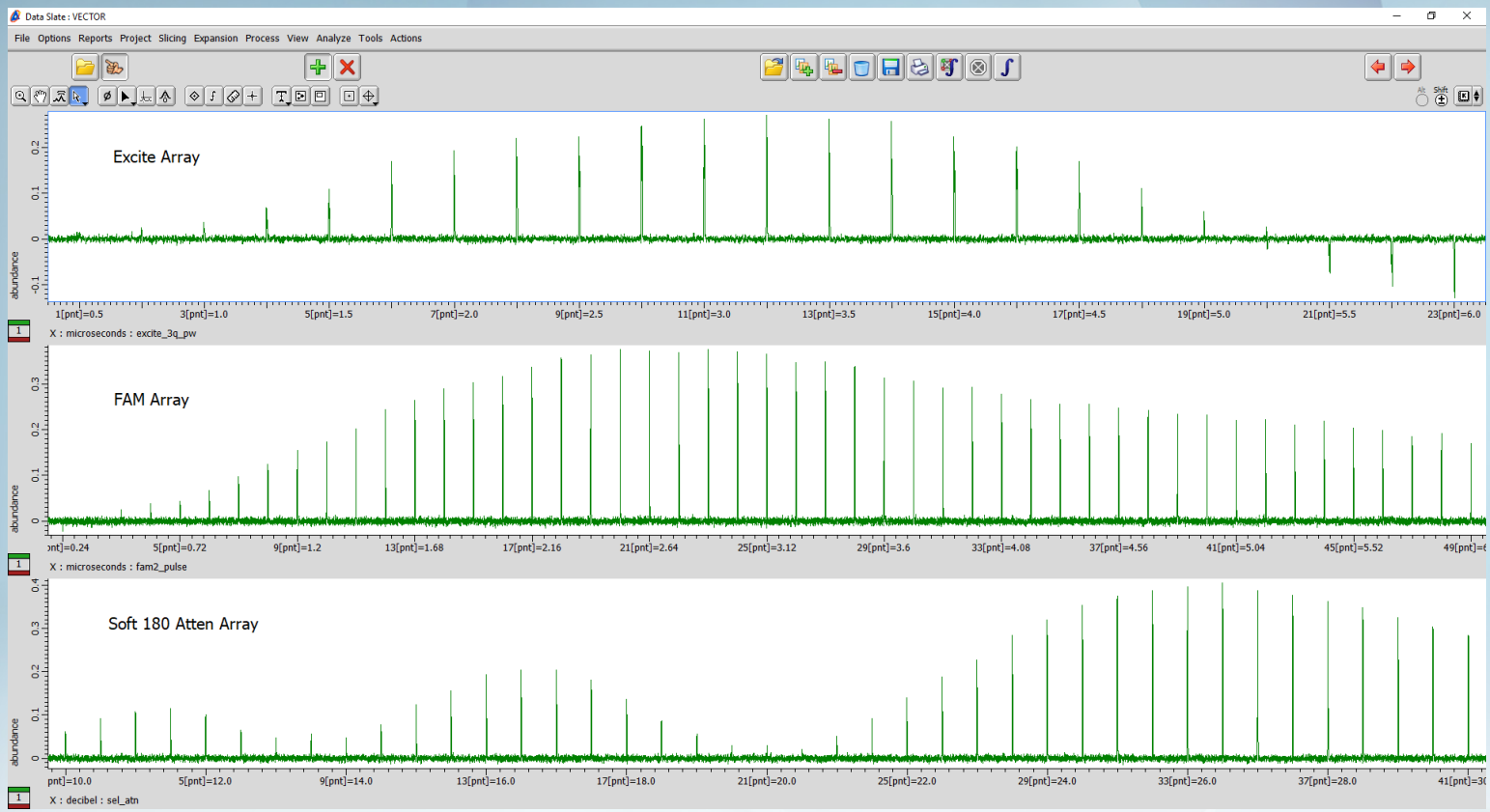


# <sup>27</sup>Al Nutation plot

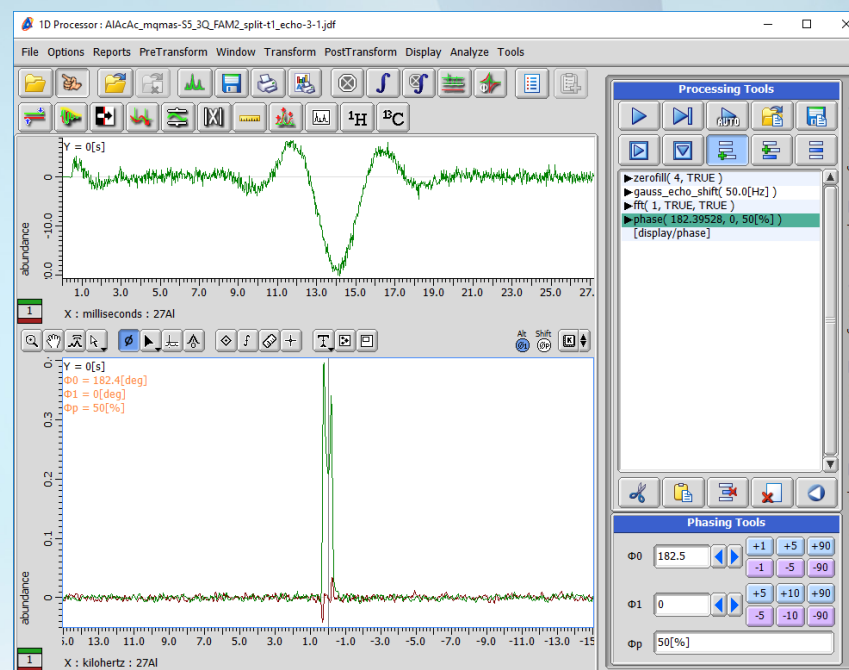
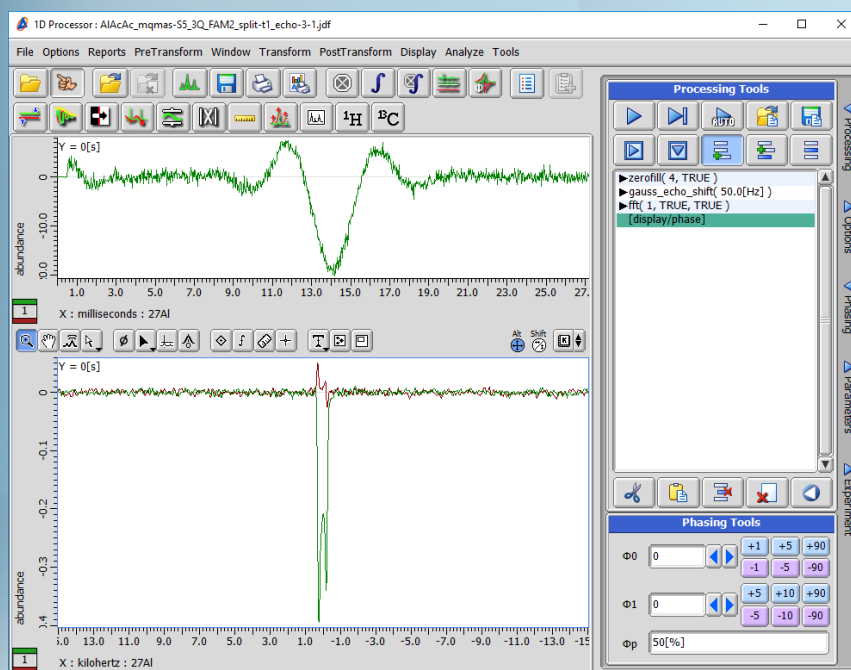


The nutation rate is scaled by the spin,  $w_n = (I + 1/2)w_{rf}$

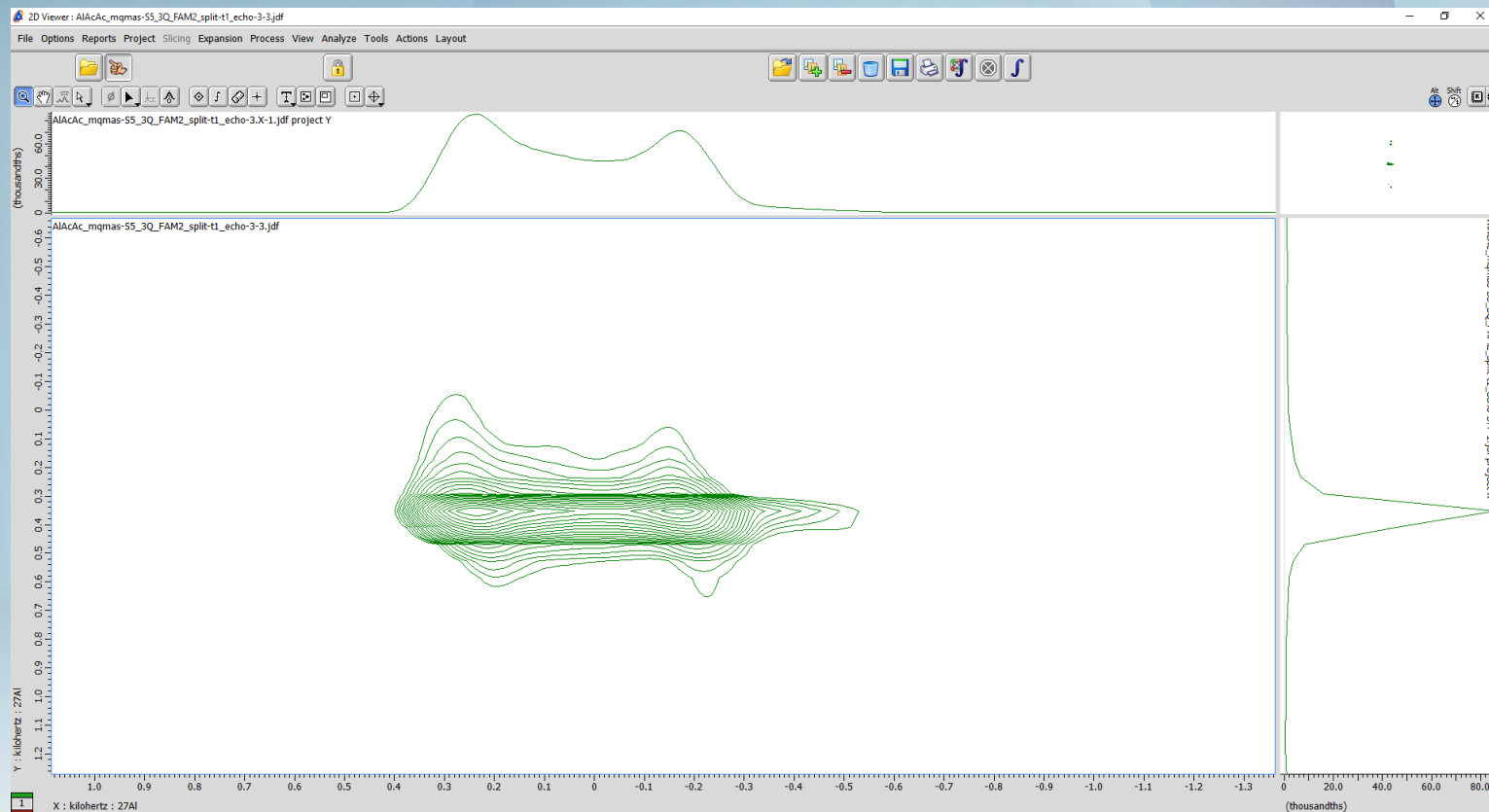
# Optimize each pulse by arraying



# Process and phase the echo

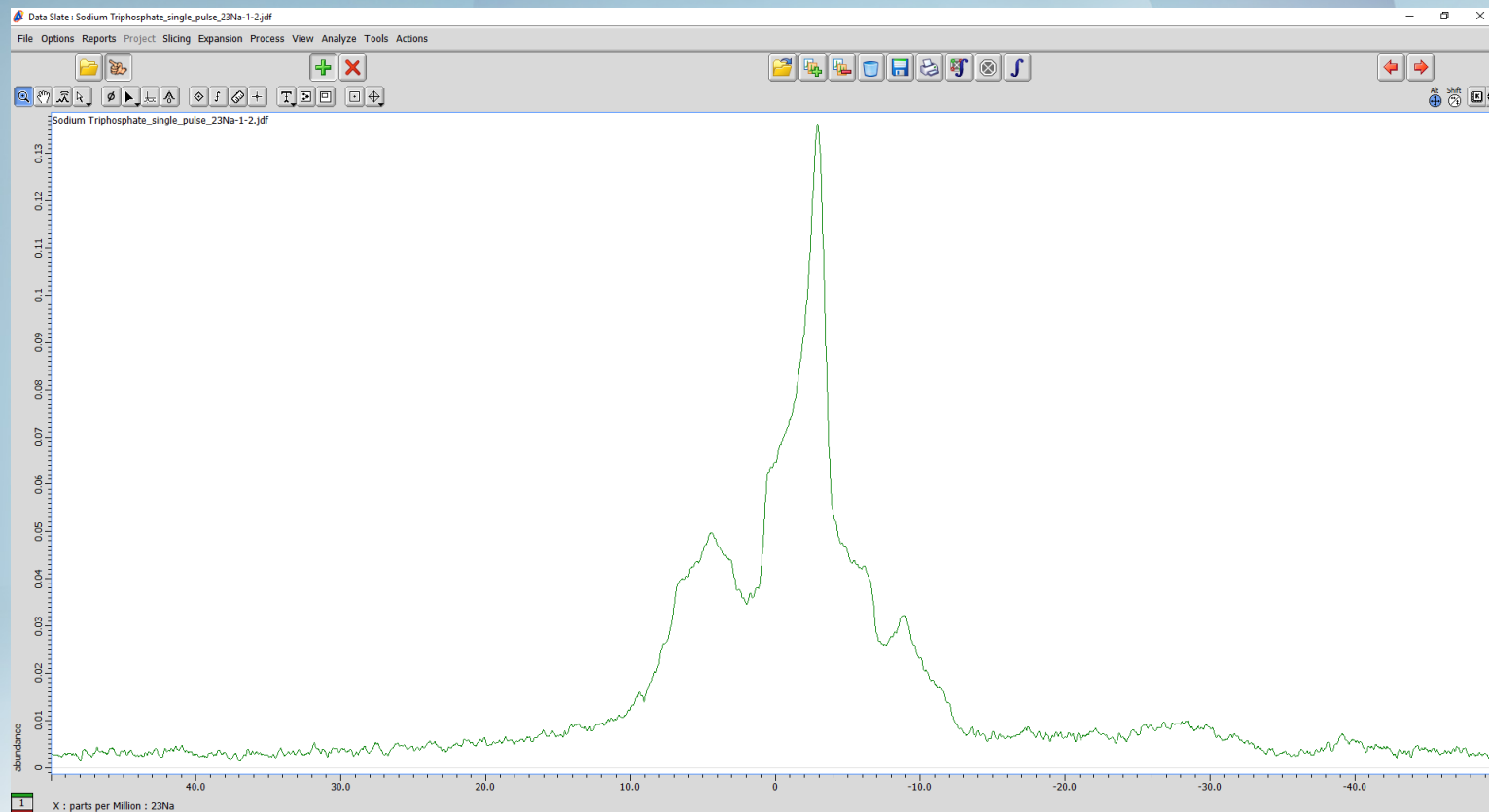


# Resulting $^{27}\text{Al}$ 3Q MQMAS of AlAcAc

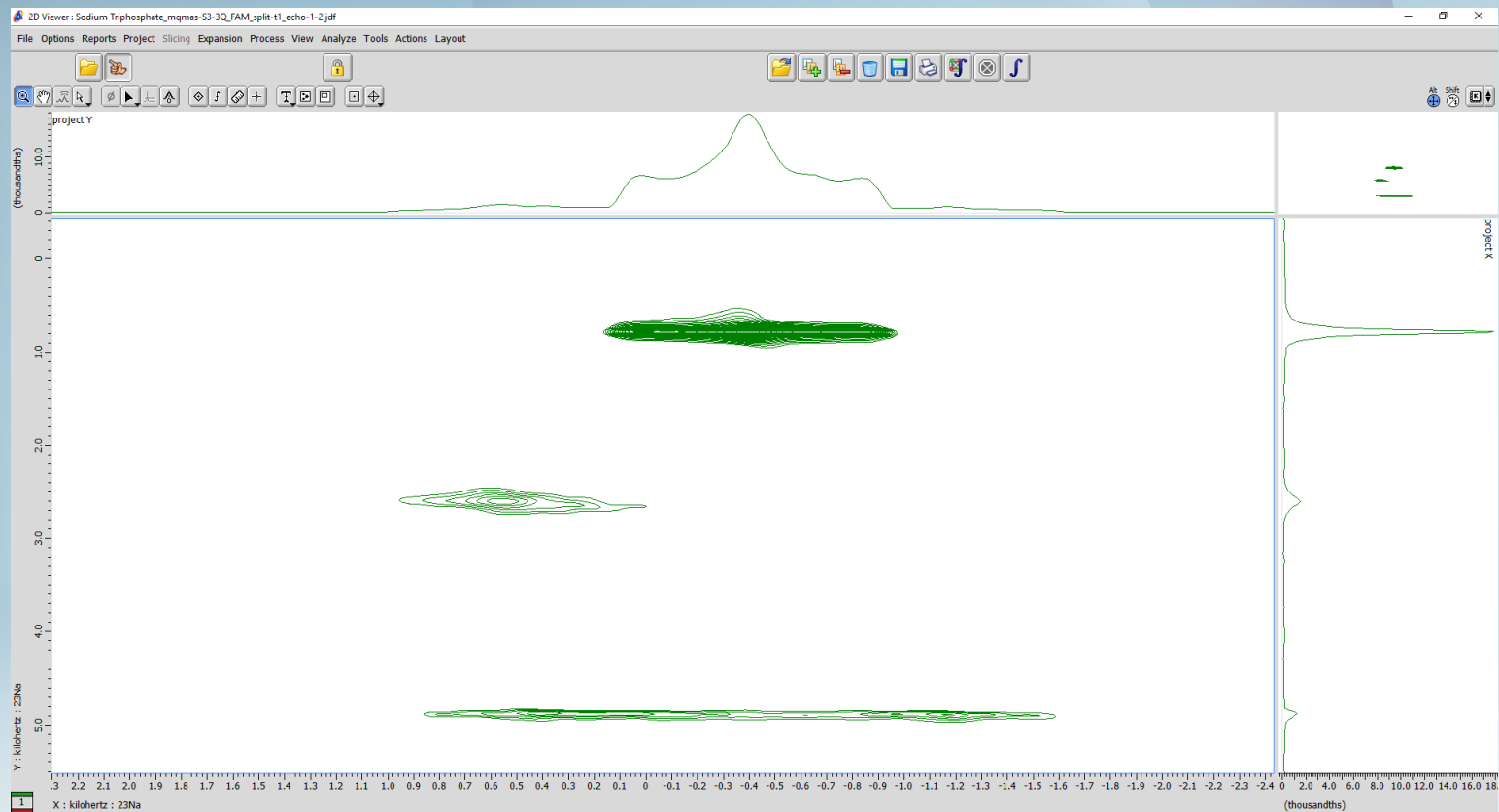




# $^{23}\text{Na}$ NMR of Sodium Triphosphate example, $\text{Na}_5\text{P}_3\text{O}_{10}$



# $^{23}\text{Na}$ NMR of Sodium Triphosphate example, $\text{Na}_5\text{P}_3\text{O}_{10}$



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    - Can destroy the probe!
  - **What nuclei do you want to observe?**
    - Is the machine capable and will the probe tune?
- **If you do not spend the time setting up properly you will get exactly what you deserve NOISE!!**